

Leaf Litter Under Changing Climate: Will Increasing Levels of CO₂ and O₃ Affect Decomposition and Nutrient Cycling Processes?

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ABSTRACT

In this manuscript we review existing information about CO₂ and O₃ effects on leaf litter as well as discuss the potential impacts of climate change on decomposition processes and tree nutrition. So far field studies show that the average response to elevated CO₂ is increased litter production and increased litter C:N-ratios, the latter response being more prominent in deciduous than in coniferous trees. The few O₃ studies indicate that O₃ stress may decrease some nutrient concentrations, but O₃ effects on carbon-based compounds are more ambiguous. In general, field incubation studies show only some small or inconsistent CO₂- and O₃-induced changes in litter mass loss rates. On the other hand, recent long-term studies indicate that there are some CO₂ and O₃ effects on microbial functioning in the soil (e.g. CO₂ stimulates and O₃ dampens it), although the onset of these microbial responses may take years. Nonetheless, at the moment there is no consistent evidence of CO₂-induced and microbe-mediated progressive nitrogen limitation in temperate forests. Elevated O₃ effects on nutrient cycling are far less studied, and also more information about long-term CO₂ and O₃ effects on decomposition and nutrient cycling in boreal forests is still needed. Furthermore, some results indicate that the combined effects cannot be predicted on the basis of single exposures and therefore, the effects of increasing CO₂ and O₃ on decomposition and nutrient cycling processes must be studied in combination.

Keywords: carbon, carbon dioxide, coniferous trees, deciduous trees, decomposition, leaf litter, lignin, microbial activity, nitrogen, ozone, phenolic compounds, soil food web

Abbreviations: **B**, boron; **C**, carbon, **Ca**, calcium; **CO₂**, carbon dioxide; **Cu**, copper; **FACE**, free-air carbon dioxide enrichment; **Fe**, iron; **K**, potassium; **LMWP**, low-molecular-weight phenolic; **Mg**, magnesium; **Mn**, manganese; **N**, nitrogen; **NH⁺₄**, ammonium; **NO_x**, nitrogen oxides; **NUE**, nitrogen use efficiency; **O₃**, ozone, **OTC**, open-top chamber; **P**, phosphorus; **PNL**, progressive nitrogen limitation; **ppb**, parts per billion, nl l⁻¹; **ppm**, parts per million, µl l⁻¹; **S**, sulphur; **SOM**, soil organic matter; **VOC**, volatile organic compound; **Zn**, zinc

CONTENTS

INTRODUCTION.....	58
EFFECTS OF ELEVATED CO ₂ ALONE	60
Litter quantity and quality responses	60
Decomposition and soil food web responses	62
EFFECTS OF ELEVATED O ₃ ALONE AND IN COMBINATION WITH CO ₂	63
Litter quantity and quality responses	63
Decomposition and soil food web responses	64
CONCLUSIONS.....	65
REFERENCES.....	66

INTRODUCTION

Before the Industrial Revolution, the atmospheric CO₂ level was around 280 ppm, whereas in 2005 the CO₂ concentrations in the surface layer of atmosphere had reached a level of 379 ppm (IPCC 2007a), which represents a global increase of 35% from the pre-industrial situation. With current climate change mitigation policies, global surface CO₂ levels will continue to grow over the next few decades, and are expected to be doubled from the pre-industrial level by the year 2100 mainly because of burning of fossil fuels and land-use change (Keeling *et al.* 1995; IPCC 2007b). Simultaneously, the annual mean concentrations of O₃ have reached levels of 20 to 50 ppb worldwide (Carter and La

Rovere 2001; Vingarzan 2004). This increase of surface levels of O₃ is expected to continue, and some scenarios suggest that the average increase of surface O₃ levels will be 8-10% at the global level during the next decade (Collins *et al.* 2000; Jonson *et al.* 2001; IPCC 2001; Vingarzan 2004). The primary reason behind the increased O₃ levels in the troposphere is the human activity and its effect on ozone precursors (i.e., O₃ is generated from NO_x and VOCs in the presence of sunlight; Fowler *et al.* 1998). In particular global emissions of NO_x have increased due to energy production and transport during the past three decades (Vingarzan 2004; Ashmore 2005).

In the near future forest ecosystems will be increasingly exposed to the joint effects of CO₂ and O₃. Although there

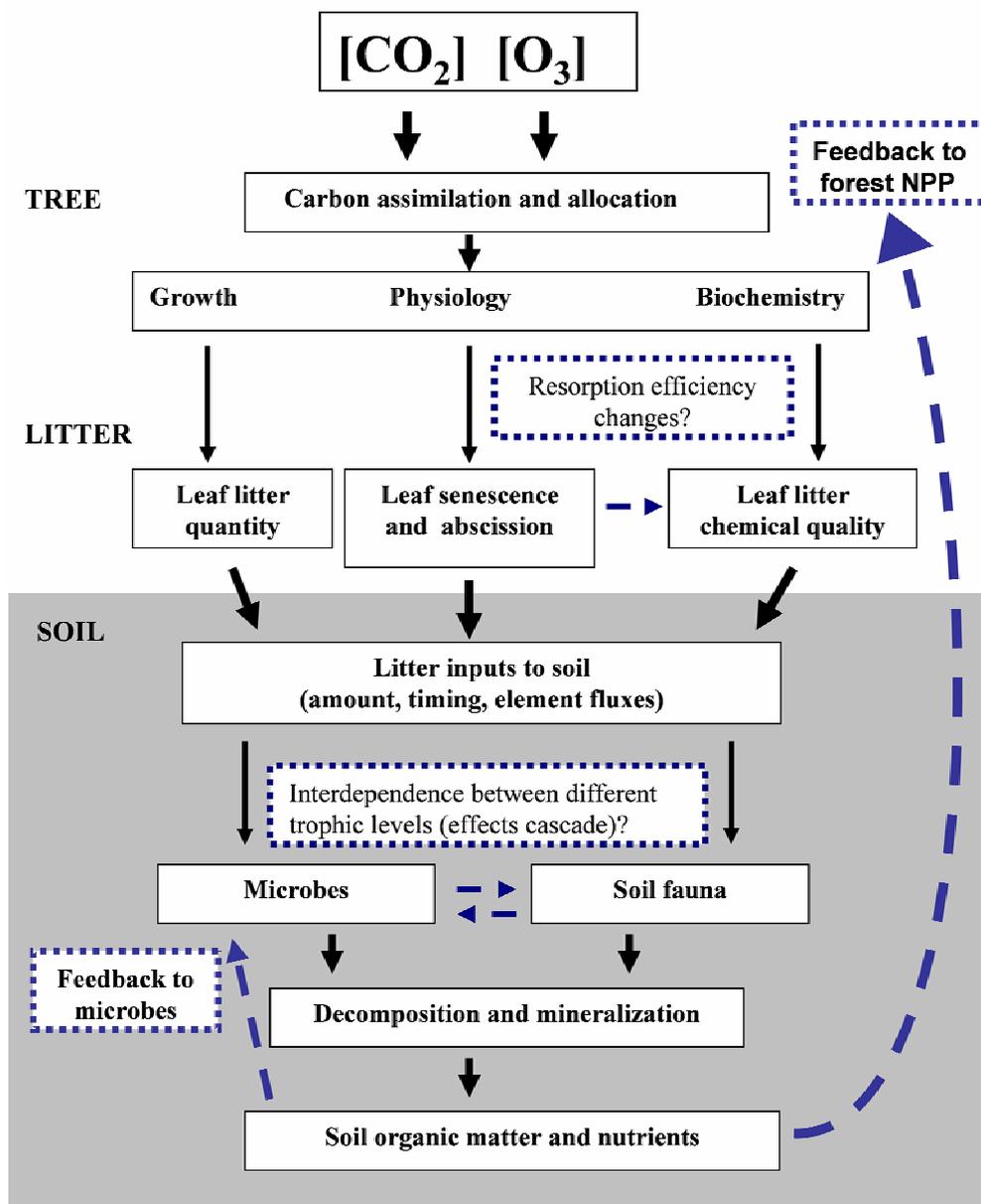


Fig. 1 Conceptual model of potential CO_2 and O_3 effects on trees and leaf litter, including links to soil food web, decomposition and mineralization processes and net primary productivity (NPP). See text for details.

are already a substantial number of CO_2 and O_3 studies about tree growth and physiology (Andersen 2003; Ainsworth and Long 2005; Grantz *et al.* 2006), the effects of these two greenhouse gases on decomposition and thereby nutrient cycling is still inadequately understood. The effects of elevated CO_2 and O_3 on decomposition are considered to be mainly indirect (Fig. 1). Hence, neither of these gases penetrates the soil extensively (Turner *et al.* 1973; Blum and Tingey 1977; Lamborg *et al.* 1983), but instead both CO_2 and O_3 first affect the tree C assimilation and C allocation within the tree, thereby influencing tree growth, physiology and biochemistry (Andersen 2003; Ainsworth and Long 2005). In theory, CO_2 - and O_3 -induced changes in green foliage are considered to be transmitted to leaf litter quantity and quality, and therefore changes in decomposition and element cycling processes as well as in net primary production (NPP) have been suggested to occur (Strain and Bazzaz 1983; Andersen 2003). However, it has been also suggested that elevated CO_2 and O_3 levels may alter leaf senescence and abscission processes (Norby *et al.* 2001; Andersen 2003) and thereby alter timing of leaf litter inputs to the soil and nutrient resorption efficiency in the foliage. If nutrient resorption efficiency changes under elevated CO_2 and O_3 , litter chemistry responses may be masked or partly

differ from those observed in green foliage. On the other hand, CO_2 and O_3 may affect decomposition via altered soil environment. For instance, CO_2 - or O_3 -induced changes in NPP can result in changed litter inputs and element fluxes to the soil or altered soil moisture conditions, and thereby affect soil biota functioning and biomass. Soil microbial responses to altered C fluxes may depend on the pre-existing soil organic matter pools and nutrient availability (Hu *et al.* 1999; Zak *et al.* 2000a, 2000b) in soil. If there are CO_2 - and O_3 -induced changes in the soil microbial biomass or community composition, these effects may also be propagated through higher levels of the soil food web, as there is a high degree of interdependence among different trophic levels (Coûteaux and Bolger 2000; Andersen 2003). Finally, although beyond the scope of this review, increasing levels of CO_2 and O_3 in the troposphere are leading to the global warming (IPCC 2007a) and in the future this climate warming may alter the decomposition processes substantially.

Since the CO_2 and O_3 effects on plant growth and physiology have been usually opposite, it has been widely suggested that elevated CO_2 may protect trees from O_3 stress (Volin *et al.* 1998; Podila *et al.* 2001). However, it is difficult to predict ecosystem-level responses because the CO_2 responsiveness and O_3 sensitivity varies widely both within

and among tree species (Karnosky *et al.* 2003). Largely based on above-ground tree growth and physiology data, responses to combined exposure are variable and can range from positive to negative or no net effect thus making predicting even more difficult. Because the above-ground responses to CO₂ and O₃ may partly differ in magnitude or direction from those of below-ground, they do not necessarily reliably describe the possible response patterns in the decomposition processes.

In this chapter we review existing literature about elevated CO₂ and O₃ effects on decomposition and nutrient cycling. We focus on leaf litter studies made with various deciduous and coniferous tree species. A majority of the leaf litter studies have been single-factorial studies, i.e., either CO₂ or O₃ studies, and mainly concentrated on leaf litter chemical quality changes and subsequent decomposition after the CO₂ and O₃ exposures. In litter studies the most commonly measured variables include N, C and lignin concentrations, C:N-ratios, and mass loss of leaf litter. However, recently there have been also some published results about leaf litter decomposition under elevated gas levels, and a few studies have included the CO₂ and O₃ effects on soil biota (microbes and soil fauna) or element (C and nutrient) cycling in the soil.

EFFECTS OF ELEVATED CO₂ ALONE

Litter quantity and quality responses

Elevated [CO₂] has usually enhanced leaf litter production in trees (Norby *et al.* 2002; Liu *et al.* 2005), but there are also results which indicate no CO₂-induced increases (Cotrufo *et al.* 2005) and species- or genotype-dependent responses to CO₂ in leaf litter production (Finzi *et al.* 2002; Riikonen *et al.* 2004). Recently Hungate *et al.* (2006) reported that in a scrub oak stand litterfall production increased initially under elevated CO₂ treatment, but after the fifth exposure season this CO₂ stimulation declined. The authors (Hungate *et al.* 2006) suggested that the declining trend was due to the accumulation of N in the surface soil organic matter, which caused the apparent restriction of plant available N and reduced the positive litter production response to CO₂ over time. On the other hand, two long-term FACE studies have shown sustained CO₂ stimulation of litter production even after six years of CO₂ exposure (Finzi *et al.* 2006; Norby and Iversen 2006). In a sweetgum stand the CO₂-induced litter production was sustained by enhanced N uptake (Norby and Iversen 2006), while in a loblolly pine dominated stand (Finzi *et al.* 2006), CO₂-exposed trees increased both their N uptake and nitrogen use efficiency and thereby sustained the increased litter production.

So far litter studies have shown either delay, enhancement or no change in leaf senescence or abscission rates (McConnaughay *et al.* 1996; Li *et al.* 2000; Johnson *et al.* 2001; Riikonen *et al.* 2004; Tricker *et al.* 2004; Cotrufo *et al.* 2005), and neither there is clear evidence of CO₂-induced changes in N resorption in trees (Norby *et al.* 2000; Finzi *et al.* 2001; Lindroth *et al.* 2001; Norby *et al.* 2001; Finzi and Schlesinger 2002; Kasurinen *et al.* 2006). Thus, the generally seen CO₂-induced decreases in N and thereby increases in C:N-ratio of green foliage should be carried over to the litter quality (Gifford *et al.* 2000; Norby *et al.* 2001). Support for this statement comes from several pot and field studies where the average CO₂ response is a decrease in N concentrations and an increase in leaf litter C:N-ratio in various tree species (Cotrufo and Ineson 1996; Cotrufo *et al.* 1998a; Gifford *et al.* 2000; Norby *et al.* 2001; **Table 1**). When we calculated the average response ratios for tree litter N across individual open-top chamber (OTC) and free-air CO₂ enrichment (FACE) experiments, the average CO₂ response for litter N was stronger (12% lower N in CO₂ treatments across the studies, **Table 1**) than that observed previously in the meta-analysis of Norby *et al.* 2001 (7% lower N in CO₂ treatments across the studies). The discrepancy between the above results can be explained

by the fact that Norby *et al.* (2001) included data also from experiments where the leaf litter N responses of non-woody species were investigated and did not separate growth chamber and pot seedling studies in the compilation, whereas we used only data from OTC- and FACE-experiments conducted with soil-growing trees.

In the meta-analysis of Norby *et al.* (2001) the N response in individual studies was rarely statistically significant, whereas in our comparison (**Table 1**) some individual studies showed clear CO₂-induced decreases in litter N concentrations (e.g. De Angelis 2000; King *et al.* 2001b; Parsons *et al.* 2004; Cotrufo *et al.* 2005; Kasurinen *et al.* 2006). However, sometimes these N reductions were observed only after the first exposure season (Norby *et al.* 2000) or were tree community-dependent (Liu *et al.* 2005). In our data compilation (**Table 1**) the average trend is increasing C:N-ratio (19% higher C:N-ratio in elevated CO₂) across various CO₂ studies with soil-growing trees. Earlier Norby *et al.* (2001) concluded that the change in C:N-ratio in soil-growing trees may not necessarily be as large and universal as previously predicted on the basis of short-term pot experiments, and also N nutrition status may modify responses to CO₂ (Gifford *et al.* 2000). For instance, King *et al.* (2001a) observed a clear CO₂-induced increase in C:N-ratios of trembling aspen litter only under low soil N availability, whereas N addition to soil removed the CO₂ effect on litter C:N-ratio. On the other hand, in another study with sugar maple trees (King *et al.* 2001b), the initial litter C:N-ratios decreased under high CO₂ + low N and increased under high CO₂+high N treatment (**Table 1**). In addition, Hättenschwiler and Bretscher (2001) observed that both the soil type and N deposition can modify litter N responses to CO₂. Thus, at the calcareous soil site high CO₂ resulted in clear decreases in beech leaf N concentration under both high and low soil N, although this negative CO₂ response was somewhat bigger in the low N treatment (**Table 1**). At the acidic site, instead, high CO₂ clearly decreased beech litter N under high N deposition only, whereas under low N deposition the CO₂ effect on N was negligible (**Table 1**). A recent meta-analysis of green foliage data also indicates that N response to CO₂ may be more prominent in deciduous than in coniferous trees (Zvereva and Kozlov 2006). In our summary (**Table 1**), all the significant leaf litter N reductions were also observed in deciduous tree species. However, there is still a lack of OTC- and FACE- experiments where naturally abscised needle litter has been investigated.

In contrast to leaf litter N concentrations (Norby *et al.* 2001; **Table 1**), litter P concentrations have generally increased under elevated CO₂. In our data compilation (**Table 1**) the average P response was +12% across CO₂ studies, but usually these CO₂-induced increases were not statistically significant in the individual experiments (**Table 1**). Finzi *et al.* (2001) observed that in a loblolly pine dominated stand the P response to CO₂ differed between the green foliage (no CO₂-induced increase in P) and leaf litter (CO₂-induced increase in P) and especially in red bud trees. However, when Finzi *et al.* (2001) calculated P resorption proficiency in red bud trees, they noticed that it was only transiently decreased due to CO₂ enrichment (i.e. CO₂ effect on P resorption was significant only after the first exposure season and then disappeared). In our study with soil-growing birch trees the P concentration response to CO₂ was similar between green foliage and leaf litter indicating that the P resorption proficiency was not significantly altered over the three-year CO₂ exposure (Kasurinen *et al.* 2006; **Table 1**). The reason for the increased P concentrations both in green foliage and leaf litter (Kasurinen *et al.* 2006) could be that in contrast to N, plant P requirement for photosynthetic machinery is not necessarily down-regulated (Conroy and Hocking 1993), but instead, the stimulation of photosynthesis may increase the P requirement for the phosphorylated photosynthetic intermediates and intercellular transport under elevated CO₂ (Gifford *et al.* 2000). Liu *et al.* (2007) also reported statistically significant increases in litter P concentrations under elevated CO₂ in aspen and aspen-birch

Table 1 Summary of CO₂ effects on leaf litter chemistry. Treatment effect % = (the CO₂ treatment average response minus the control treatment average response divided by the control treatment average response) × 100. Negative values indicate decrease and positive values increase in chemical concentration or C:N-ratios, and 0 denotes for no change. Bolded values indicate an average response to CO₂ treatment across the individual experiments. Symbols: * = a significant CO₂ effect (P ≤ 0.1), ** = tree community-dependent CO₂ response, (*) = CO₂ effect transient. Only data from OTC- and FACE-studies with soil-growing trees and naturally abscised leaf litter was included. Abbreviation nd = not determined.

Tree species/community	N	P	C:N-ratio	Lignin	Condensed tannins	Reference
<i>Acer rubrum</i> (ambient temperature)	-18 ^(*)	nd	nd	nd	nd	Norby <i>et al.</i> 2000 ¹
<i>Acer rubrum</i> (elevated temperature)	-10 ^(*)	nd	nd	nd	nd	Norby <i>et al.</i> 2000 ¹
<i>A. rubrum</i>	-8	+26	nd	-3	nd	Finzi <i>et al.</i> 2001 ¹
<i>A. rubrum</i>	-4	nd	nd	+2	nd	Finzi and Schlesinger 2002 ¹
<i>A. saccharum</i> (ambient temperature)	-7 ^(*)	nd	nd	nd	nd	Norby <i>et al.</i> 2000 ¹
<i>A. saccharum</i> (elevated temperature)	-12 ^(*)	nd	nd	nd	nd	Norby <i>et al.</i> 2000 ¹
<i>A. saccharum</i> (low N)	-10*	nd	-7*	nd	+31*	King <i>et al.</i> 2001b
<i>A. saccharum</i> (high N)	-18*	nd	+26*	nd	+40*	King <i>et al.</i> 2001b
<i>Betula papyrifera</i>	-31*	nd	+44*	+5	+64*	Parsons <i>et al.</i> 2004
<i>B. pendula</i>	-11*	+18*	+12*	+4*	+23*	Kasurinen <i>et al.</i> 2006, 2007 ²
<i>B. papyrifera</i> - <i>Populus tremuloides</i>	-18*	+14*	+20*	+6	+63**	Liu <i>et al.</i> 2005, 2007
<i>Cornus florida</i>	-26	+6	nd	-19	nd	Finzi <i>et al.</i> 2001 ¹
<i>C. florida</i>	-13	nd	nd	0	nd	Finzi and Schlesinger 2002 ¹
<i>Cercis canadensis</i>	0	+38	nd	-6	nd	Finzi <i>et al.</i> 2001 ¹
<i>C. canadensis</i>	-14	nd	nd	+1	nd	Finzi and Schlesinger 2002 ¹
<i>Fagus sylvatica</i> (acidic soil+low N)	-2	nd	nd	+3	nd	Hättenschwiler and Bretscher 2001
<i>F. sylvatica</i> (calcareous soil+low N)	-21*	nd	nd	+4	nd	Hättenschwiler and Bretscher 2001
<i>Fagus sylvatica</i> (acidic soil+high N)	-15*	nd	nd	+8*	nd	Hättenschwiler and Bretscher 2001
<i>F. sylvatica</i> (calcareous soil+high N)	-11*	nd	nd	0	nd	Hättenschwiler and Bretscher 2001
<i>Liquidambar styraciflua</i>	-8	+19	nd	+8	nd	Finzi <i>et al.</i> 2001 ¹
<i>L. styraciflua</i>	-15	nd	nd	+3	nd	Finzi and Schlesinger 2002 ¹
<i>Pinus taeda</i>	+3	+16	nd	-1	nd	Finzi <i>et al.</i> 2001 ¹
<i>P. taeda</i>	0	nd	nd	+5	nd	Finzi and Schlesinger 2002 ¹
<i>P. sylvestris</i>	-7	-11	nd	nd	nd	Kainulainen <i>et al.</i> 2003
<i>Populus alba</i>	-5*	nd	+7*	-7	nd	Cotrufo <i>et al.</i> 2005
<i>P. × euramericana</i>	-36*	nd	+45*	-4	nd	Cotrufo <i>et al.</i> 2005
<i>P. nigra</i>	-23*	nd	+29*	+2	nd	Cotrufo <i>et al.</i> 2005
<i>P. tremuloides</i>	-14*	+5*	+13*	+37	-18**	Liu <i>et al.</i> 2005, 2007
<i>Quercus ilex</i>	-12*	-13	+21*	+19*	nd	de Angelis <i>et al.</i> 2000 ¹
<i>Q. myrtifolia</i> (Exp. 1)	0	nd	+1	-8	+12	Hall <i>et al.</i> 2006
<i>Q. myrtifolia</i> (Exp. 2)	-2	nd	+2	+30*	+2	Hall <i>et al.</i> 2006
Average response (n = 8 - 31)	-12	+12	+18	+4	+27	

communities. They (Liu *et al.* 2007) suggested that the CO₂-induced increase in P concentrations of leaf litter could be due to a concurrent increase in cuticular wax production (Percy *et al.* 2002), which could have reduced P leaching from the leaves under elevated CO₂.

The impact of CO₂ enrichment on other nutrients than N and P are far less studied. Cotrufo *et al.* (1998a) reviewed pot seedling and growth chamber studies and did not find any clear CO₂ responses in tree litter K, Ca, Mg, Mn and Fe concentrations. The few field experiments with soil-growing trees have not either been able to provide evidence of strong or consistent CO₂ effects on other nutrients. For instance, Kainulainen *et al.* (2003) did not find any CO₂ treatment effect on K, Ca and Mg concentrations of Scots pine needle litter after a three-year CO₂ exposure. In our study with two silver birch clones S concentration decreased under elevated CO₂ levels, but the other analysed nutrients (Ca, Mg, Mn, Fe, Zn, Cu, B) did not show any consistent change or the CO₂-induced changes were largely genotype-dependent (e.g. decrease in K concentrations) (Kasurinen *et al.* 2006). Liu *et al.* (2007) observed that elevated CO₂ marginally significantly increased litter K and S concentrations and significantly decreased B concentrations in both aspen and aspen-birch communities while litter Mn responses to CO₂ were community-dependent.

In addition to CO₂-induced changes in the amount of total C entering the soil, the changes in litter C compound composition (i.e. changes in C quality) may also play a significant role in the subsequent decomposition processes (Horner *et al.* 1988; Zak *et al.* 1993; Hättenschwiler and Vitousek 2000; Zak *et al.* 2000a, 2000b). The carbon-nutrient hypothesis (Bryant *et al.* 1983) and its extension, the growth-differentiation balance hypothesis (Herms and Matt-

son 1992), predict that changes in source-sink relationship could lead to variations in the relative partitioning of carbon to growth, total non-structural carbohydrates and carbon-based secondary and structural compounds. Thus, under elevated [CO₂] carbon source may be higher than carbon sink, and excess carbon could lead to the over-investment of non-structural carbohydrates and secondary or structural C compounds (Koricheva *et al.* 1998; Peñuelas and Estiarte 1998; Peltonen *et al.* 2005), especially under low soil N conditions (Lambers 1993). However, the magnitude of this CO₂ response has been observed to vary between different exposure systems (Norby *et al.* 2001) and C compound groups (Zvereva and Kozlov 2006). For instance, in pot seedling studies leaf litter lignin concentrations usually clearly increased under elevated CO₂, but OTC- or FACE-studies with soil-growing trees do not uniformly support these early findings (Norby *et al.* 2001, **Table 1**). In fact, only a few OTC studies have reported statistically significant increases in lignin concentrations under elevated CO₂ (DeAngelis *et al.* 2000; Hall *et al.* 2006; Kasurinen *et al.* 2006). When we summarized condensed tannin data across the OTC- and FACE-experiments (only tree litter results were used), we observed that in contrast to lignin, the average CO₂ response was relatively high (27% increase on average) and that in most individual studies this CO₂ effect was also statistically significant (**Table 1**). Furthermore, our OTC experiment indicated that elevated CO₂ has the potential to increase total phenolics and cellulose concentrations in silver birch litter (Kasurinen *et al.* 2006, 2007), while other studies with various tree species show no CO₂-induced increases in phenolics (Kainulainen *et al.* 2003) or cellulose concentrations (Liu *et al.* 2005; Hall *et al.* 2006). Of less studied terpene compounds, α- and β-pinene has

been observed to increase due to CO₂ enrichment, whereas total monoterpenes have not shown any clear CO₂-induced increase in Scots pine litter (Kainulainen *et al.* 2003).

Decomposition and soil food web responses

Results to date from litter decomposition studies have been variable and inconclusive (O'Neill and Norby 1996; Cotrufo *et al.* 1998a, 1998b; Coûteaux *et al.* 1999; Norby *et al.* 2001; **Table 2**). Laboratory studies with CO₂-exposed material have shown substantial rate retarding effects of CO₂ on the subsequent litter decomposition, but field litter incubations usually have not been able to confirm these early findings (O'Neill and Norby 1996; Norby *et al.* 2001). Thus, with few exceptions, a majority of the field incubation studies conducted under ambient CO₂ conditions have not detected any significant or consistent CO₂-induced changes in the decomposition rates (O'Neill and Norby 1996; Norby *et al.* 2001; **Table 2**). In our three-year study with silver birch, the negative CO₂ effect on decomposition was genotype-dependent and transient as elevated CO₂ decreased the subsequent decomposition in one birch clone only after the first and second exposure season (Kasurinen *et al.* 2006). On the other hand, some field incubation studies indicate that the CO₂ environment itself may modify the leaf litter decomposition responses significantly (**Table 2**). Parsons *et al.* (2004) reported that under ambient CO₂ plots, paper birch leaf litter produced in high CO₂ treatments decomposed slower than that produced in ambient CO₂ treatments, whereas in elevated CO₂ plots, the CO₂ enrichment itself during the decomposition process reinforced the negative effects of CO₂ on mass loss even further. Hall *et al.* (2006) observed that high CO₂ environment itself enhanced scrub oak litter decomposition, but the litter origin (i.e. whether it was produced under ambient or elevated CO₂ level) did not affect the decomposition rates in ambient or elevated CO₂ environment. Cotrufo *et al.* (2005) found that independent of the poplar species, litter generated under elevated CO₂ had slightly lower mass loss rates than litter produced under ambient CO₂, whereas in high CO₂ environment the decomposition rates of leaf litter of two poplar species were enhanced regardless of the litter origin (i.e. whether it was from ambient or elevated CO₂ plot). Thus, basically Cotrufo *et al.* (2005) observed that under

ambient CO₂ environment CO₂-induced changes in leaf litter quality governed the decomposition rates, whereas in elevated CO₂ plots the CO₂-induced changes in soil environment itself may have controlled the litter disappearance. However, the authors (Cotrufo *et al.* 2005) did not give any specific causative mechanism for the latter observation, e.g., whether the increased decomposition in CO₂ plots was due to enhanced microbial activity or some other factor. In a loblolly pine-dominated stand, leaf litter decomposition of five different tree species (**Table 2**) was not significantly affected by the CO₂ exposure during the litter decomposition (Finzi *et al.* 2001; Finzi and Schlesinger 2002).

The role of soil animals in the actual decomposition process is secondary compared to soil microbes in temperate and boreal vegetation zones (Berg and Laskowski 2006). However, soil fauna can have an important role in soil formation and nutrient cycling by mixing the leaf litter into deeper soil layers and releasing nutrients by grazing both the leaf litter and litter-associated microbes (Lavelle and Spain 2001; Hättenschwiler and Gasser 2005). Soil fauna also interacts with soil and litter-associated microbes meaning that animals and microbes can stimulate each other's activity during the decomposition process (Coûteaux *et al.* 1991, 1996; Lavelle and Spain 2001; Hättenschwiler and Gasser 2005). At present, information about litter-mediated CO₂ effects on litter-feeding soil animals is scarce, inconsistent, and mainly limited to woodlice (i.e. terrestrial isopods, important detritivores in the temperate forests). For instance, leaf litter consumption by woodlice has been observed to decrease (Hättenschwiler *et al.* 1999), increase (Cotrufo *et al.* 1998b) or not to change (Hättenschwiler and Bretscher 2001), although all of these studies have reported significant leaf litter quality changes (e.g. clear decrease in N) due to elevated CO₂. In our microcosm study (Kasurinen *et al.* 2007), the leaf litter consumption rates by woodlice were poorly related to the measured chemical components, and only a marginally significant and transient decrease in consumption rates were observed when animals were fed with N-poor and phenolic-rich CO₂ litter.

The complexity of the soil food web itself can be an important factor for modifying the CO₂ effects on the subsequent decomposition (Coûteaux *et al.* 1991; Coûteaux and Bolger 2000). For instance, the decomposition rates of CO₂-enriched sweet chestnut litter exposed to simple food webs

Table 2 Summary of CO₂ effects on subsequent leaf litter decomposition (absolute mass loss). Symbols: ↔ = no change, ↓ = decrease, (↓) = slight decrease, ↑ = increase, * = CO₂ environment enhances the negative litter-mediated CO₂ effect, ** = genotype-dependent CO₂ response, (s) = CO₂ environment effect on mass loss same regardless of the litter origin. Only data from OTC- and FACE-studies with soil-growing trees and naturally abscised leaf litter was included. Abbreviation: nd = not determined.

Tree species/community	Mass loss		Reference
	Decomposition environment		
	Ambient CO ₂	Elevated CO ₂	
<i>Acer rubrum</i>	↔	↔	Finzi <i>et al.</i> 2001
<i>A. rubrum</i>	↔	↔	Finzi and Schlesinger 2002
<i>Betula papyrifera</i> (Native placement + Common garden)	↓	↓*	Parsons <i>et al.</i> 2004
<i>B. papyrifera</i> (Common substrate)	↔	↔	Parsons <i>et al.</i> 2004
<i>B. pendula</i> (Exp. 1)	↓**	nd	Kasurinen <i>et al.</i> 2006
<i>B. pendula</i> (Exp. 2)	↓**	nd	Kasurinen <i>et al.</i> 2006
<i>B. pendula</i> (Exp. 3)	↔	nd	Kasurinen <i>et al.</i> 2006
<i>Cornus florida</i>	↔	↔	Finzi <i>et al.</i> 2001
<i>C. florida</i>	↔	↔	Finzi and Schlesinger 2002
<i>Cercis canadensis</i>	↔	↔	Finzi <i>et al.</i> 2001
<i>C. canadensis</i>	↔	↔	Finzi and Schlesinger 2002
<i>Liquidambar styraciflua</i>	↔	↔	Finzi <i>et al.</i> 2001
<i>L. styraciflua</i>	↔	↔	Finzi and Schlesinger 2002
<i>Pinus taeda</i>	↔	↔	Finzi <i>et al.</i> 2001
<i>P. taeda</i>	↔	↔	Finzi and Schlesinger 2002
<i>P. sylvestris</i>	↔	nd	Kainulainen <i>et al.</i> 2003
<i>Populus alba</i>	(↓)	↔	Cotrufo <i>et al.</i> 2005
<i>P. × euramericana</i>	(↓)	↑	Cotrufo <i>et al.</i> 2005
<i>P. nigra</i>	(↓)	↑	Cotrufo <i>et al.</i> 2005
<i>Quercus ilex</i>	nd	↓	de Angelis <i>et al.</i> 2000
<i>Q. myrtifolia</i> (Exp. 1)	↔	nd	Hall <i>et al.</i> 2006
<i>Q. myrtifolia</i> (Exp. 2)	↔	↑(*)	Hall <i>et al.</i> 2006

(microflora+Protozoa) were lower than that of control litter, whereas exposure to more complex foodwebs (microflora + Protozoa + higher soil fauna including nematodes, springtails and woodlice) led to higher decomposition rates of the CO₂-enriched litter when compared to control litter (Coûteaux *et al.* 1991; Coûteaux and Bolger 2000). In a recent FACE study with paper birch, trembling aspen and sugar maple, elevated CO₂ increased the total tree productivity but simultaneously decreased the abundance of total soil fauna, springtails and mites after a four-year-exposure period (Loranger *et al.* 2004). At the same time, Parsons *et al.* (2004) observed a decrease in the subsequent decomposition of birch leaves in the same CO₂ plots. Both results (Loranger *et al.* 2004; Parsons *et al.* 2004) indicate that the CO₂-induced changes in litter quality of both above- and below-ground compartments can be manifested in the soil food web and in its functioning in the deciduous tree stands. In a FACE study with loblolly pine trees, Hansen *et al.* (2001) noticed a consistent trend toward lower microarthropod abundance in elevated CO₂, but in this case the driving mechanism was suggested to be some change in the microbial resource base or habitat (i.e., non-litter CO₂ effects), as this negative animal response was observed early after the onset of the CO₂ treatment and before any CO₂-induced quality changes in leaf litter was even possible to detect. In contrast, Haimi *et al.* (2005) found only minor CO₂-induced changes in decomposer fauna at the Scots pine stand after six years of CO₂ exposure (i.e., densities of one mite suborder and one springtail species decreased), but since they did not study litter quality it is not possible to state whether there was also a lack of litter response to elevated CO₂.

The few available studies indicate that CO₂-induced decreases in litter N concentrations may be maintained at the initial stages of decomposition (Coûteaux *et al.* 1991, 1996; Parsons *et al.* 2004; Cotrufo *et al.* 2005). Although the lower N concentrations at the early stages of decomposition may decrease decomposition rates, the effect of N concentration is reversed during the later stages of decomposition (Berg and Laskowski 2006). Thus, the CO₂-induced changes in N concentrations of leaf litter could in fact lead to more complete decomposition (i.e., humus formation decreases), although it must be kept in mind that N is not the only nutrient controlling the litter disappearance rates at the later stages of decomposition (Berg and Laskowski 2006). For instance, Coûteaux *et al.* (1991) showed in their microcosm study that the early stage of decomposition of CO₂-enriched sweet chestnut litter was always lower than that of more N-rich control litter, but during the later stages the decomposition rate of N-poor and CO₂-exposed litter in fact enhanced. In another incubation study with sweet chestnut litter (Coûteaux *et al.* 1996), lignin decay started earlier in N-poor CO₂ litter than in more N-rich control litter, whereas Parsons *et al.* (2004) did not observe any significant change in lignin decomposition rates of birch litter both produced and incubated under elevated CO₂. Although a laboratory study (King *et al.* 2001b) indicated that initial CO₂ treatment effects on condensed tannins (i.e. CO₂-induced increase) in sugar maple leaf litter may quickly disappear, a longer field incubation experiment in high CO₂ environment (Parsons *et al.* 2004) indicated that the initial CO₂-induced increases in the concentrations of condensed tannins in paper birch leaves are not necessarily immediately neutralized by microbes or physical effects such as leaching. Elevated CO₂ has not been found to affect the subsequent disappearance rates of condensed tannins in trembling aspen leaves (King *et al.* 2001a) or total phenolics and monoterpenes in Scots pine needles (Kainulainen *et al.* 2003).

According to the concept of progressive N limitation, declining N mineralization (i.e. microbial N release from litter and SOM) is one of the indicators of incipient PNL in N-limited ecosystems (Luo *et al.* 2004). So far the results of CO₂ enrichment experiments have been ambiguous and there is no clear support for either positive (Zak *et al.* 1993) or negative feedback (Diaz *et al.* 1993) on soil N cycling as

various studies have reported either increases, decreases or no change in the rate of N immobilization and mineralization under elevated CO₂ (Finzi and Schlesinger 2003; Holmes *et al.* 2003; Zak *et al.* 2003; Cotrufo *et al.* 2005; Holmes *et al.* 2006). However, the CO₂-response of microbial functioning may change over time, as Holmes *et al.* (2003) first observed no change in N transformations beneath CO₂-exposed trees, but later they measured increased gross N mineralization rates and NH₄⁺ immobilization from the same experimental plots (Holmes *et al.* 2006). In the same FACE experiment with trembling aspen, paper birch and sugar maple also soil microbial activity involved with the degradation of plant cell wall components was stimulated (Larson *et al.* 2002; Phillips *et al.* 2002; Chung *et al.* 2006). Hence, N turnover by microbes was increased, but there was no proof of altered soil N cycling (Holmes *et al.* 2006). In a loblolly pine dominated forest microbial-N immobilization was not increased under elevated CO₂ and although the rate of net N mineralization declined over the first six years of experiment, this decline was not significantly more rapid under elevated CO₂ (Finzi *et al.* 2006). Previously Zak *et al.* (2000a) concluded that only in systems where the CO₂-induced increase in plant litter production clearly overcomes the pre-existing SOM pools, soil N cycling may be changed. In some ecosystems the direction and magnitude of N and C cycling response may depend more on changes in below-ground litter inputs than in above-ground litter inputs (Zak *et al.* 2000b; Matamala *et al.* 2003; de Graaff *et al.* 2006). Taken together, although some decomposition and element cycling processes in soil may be altered due to elevated CO₂, at the moment there is no consistent evidence of CO₂-induced and microbe-mediated PNL in temperate forests. Thus, more long-term studies (>7 years) including both above- and below-ground litter sources and with more complex soil food webs are needed to get an overall picture of the decomposition and element cycling processes.

EFFECTS OF ELEVATED O₃ ALONE AND IN COMBINATION WITH CO₂

Litter quantity and quality responses

Since the average effect of O₃ on tree growth is negative (Grantz *et al.* 2006), it can be assumed that in the future O₃ will decrease the leaf litter inputs to the soil, if some other environmental factor (e.g. elevated CO₂) does not counteract this negative effect. There are only a few O₃ and CO₂ × O₃ interaction studies where the actual leaf litter production has been reported. In most recent studies, Liu *et al.* (2005) reported clear O₃-induced decreases in leaf litter masses in the FACE experiment with trembling aspen and paper birch trees, whereas in our OTC study no clear O₃ effects on silver birch leaf litter mass were found over a three-year-exposure period (Riikonen *et al.* 2004). In the above FACE study, the negative O₃ effect on leaf litter production was detected mainly under ambient CO₂ levels, although elevated O₃ had also a tendency to diminish the positive effect of CO₂ on leaf production in the combination treatment (Liu *et al.* 2005). O₃ stress has also been observed to accelerate foliar senescence and abscission (Findlay *et al.* 1996; Andersen 2001, 2003; Riikonen *et al.* 2004), but based on scarce data it seems that this O₃ effect is mainly observed under ambient CO₂ conditions (Riikonen *et al.* 2004).

If leaf senescence is accelerated due to O₃ stress and N resorption from leaves is not complete at the time of leaf abscission this could lead to leaf litter with increased N levels (Findlay and Jones 1990; Uddling *et al.* 2005). So far O₃ effects on N resorption or leaf litter N have been highly variable (Scherzer *et al.* 1998; Lindroth *et al.* 2001; Uddling *et al.* 2005). Previously Lindroth *et al.* (2001) have also reported that the elevated O₃ responses were modified by the tree species and prevailing CO₂ levels. First of all, O₃ stress decreased both the N resorption efficiency and C:N-ratios in paper birch leaves, but not in trembling aspen, and secondly, under the combination treatment, the negative O₃

Table 3 Summary of O₃ effects on leaf litter chemistry. Treatment effect % = (the O₃ treatment average response minus the control treatment average response divided by the control treatment average response) × 100. Negative values indicate decrease and positive values increase in chemical concentration or C:N-ratios, and 0 denotes for no change. Symbols: * = a significant O₃ effect (P ≤ 0.1) and ** = tree community-dependent O₃ response. Only data from OTC- and FACE-studies with soil-growing trees and naturally abscised leaf litter was included. Abbreviation: nd = not determined.

Tree species/community	N	P	C:N-ratio	Lignin	Condensed tannins	Reference
<i>Betula papyrifera</i>	0	nd	+3	-12	+4	Parsons <i>et al.</i> 2004
<i>B. pendula</i>	-6	-15*	+8	+1	+5	Kasurinen <i>et al.</i> 2006, 2007 ¹
<i>B. papyrifera</i> - <i>Populus tremuloides</i>	-18*	-27*	+23**	-5	+94*	Liu <i>et al.</i> 2005, 2007
<i>Pinus sylvestris</i>	-2	-3	nd	nd	nd	Kainulainen <i>et al.</i> 2003
<i>Populus tremuloides</i>	-7*	-19*	+7**	+26	+79*	Liu <i>et al.</i> 2005, 2007
Average response (n =4-5)	-7	-16	+10	+3	+46	

¹ data averaged over three experiment years

effect on N resorption efficiency in birch leaves was cancelled due to elevated CO₂, whereas the C:N-ratios of birch leaves were increased even more than in the single CO₂ treatment (Lindroth *et al.* 2001). In our small O₃ data compilation with soil-growing trees and naturally abscised litter (Table 3), only one study (Liu *et al.* 2005) reported marginally significant O₃ effects on aspen and aspen-birch litter N concentrations (decrease) and C:N-ratios (increase) while other studies showed no clear O₃ response patterns in litter N and C:N-ratios. In addition, Kasurinen *et al.* (2006) or Parsons *et al.* (2004) did not either observe any significant CO₂ × O₃ effects on leaf N concentrations or C:N-ratios in two different birch species. In fact, in both studies the negative CO₂ effect on leaf litter N concentrations and C:N-ratios were seen regardless of prevailing O₃ levels, as elevated CO₂ decreased N concentrations and increased C:N-ratios in the CO₂+O₃ treatment also (Parsons *et al.* 2004; Kasurinen *et al.* 2006). Although in the study of Liu *et al.* (2005) both elevated CO₂ alone and O₃ alone decreased litter N concentrations, the combination treatment did not show additive response (e.g. either CO₂ or O₃ did not exacerbate each other's influence).

Table 3 shows that all the significant O₃-induced decreases in litter P concentrations have been observed in birch or aspen experiments. In both studies the O₃-induced decrease in P concentrations was observed mainly under ambient CO₂ (Kasurinen *et al.* 2006; Liu *et al.* 2007). In addition, Kasurinen *et al.* (2006) observed that in silver birch O₃ reduced the concentrations of Mn, Zn and B in both green foliage and leaf litter, and that these O₃-induced decreases in nutrients were detected also under elevated CO₂ (Kasurinen *et al.* 2006). Since the leaf litter mass was not significantly altered by O₃ treatments (Riikonen *et al.* 2004), fluxes of some of these nutrients to the soil (e.g. B and Mn) were somewhat decreased (Kasurinen *et al.*, personal data). Liu *et al.* (2007) reported O₃-induced decreases in aspen and aspen-birch litter S, Ca and Zn concentrations. In contrast to our study (Kasurinen *et al.* 2006), the negative O₃ effect on Zn was seen only under ambient CO₂ (Liu *et al.* 2007). Because of decreased litter production, elevated O₃ decreased statistically significantly the fluxes of N, P, S, K, Ca, Mg, Cu, and Zn, and marginally statistically significantly the flux of Mn to the soil (Liu *et al.* 2007). Based on this limited data, it seems that O₃ is capable of inducing some changes in leaf litter nutrient composition, but these responses are less consistent than those induced by elevated CO₂, and under combined exposure situation elevated CO₂ may either dominate over (Parsons *et al.* 2004; Kasurinen *et al.* 2006) or totally cancel the O₃ responses (Lindroth *et al.* 2001; Liu *et al.* 2007).

Ozone is a phytotoxic gas, and under O₃ stress trees may reduce their C assimilation and photosynthate production. On the other hand, O₃ is known to activate phenylpropanoid metabolism regardless of the potentially decreased photosynthesis (Kangasjärvi *et al.* 1994; Koricheva *et al.* 1998; Dizengremel 2001; Podila *et al.* 2001). Although some greenhouse studies have shown O₃-induced increases in leaf litter total phenolics (Findlay *et al.* 1996), long-term field experiments have not usually revealed any clear O₃-

induced changes in phenolics (Kainulainen *et al.* 2003; Kasurinen *et al.* 2007; but see Liu *et al.* 2005). The discrepancy between the greenhouse and field studies is probably partly due to the differences in the O₃ exposures used. For instance, Findlay and Jones (1990) used high levels of O₃ for a short exposure period (i.e. O₃ level of 200 ppb for 5 hours), whereas in recent field studies O₃ exposure levels have been usually only about two times the ambient levels (e.g., approximately 60 ppb) and exposure has continued for several consecutive growing seasons (Kainulainen *et al.* 2003; Kasurinen *et al.* 2007). However, in our OTC study with birch some O₃ effects on individual phenolic compound groups manifested gradually towards the end of the experiment (Kasurinen *et al.* 2007), this result thus emphasizing a need for long-term (3 years or more) studies in order to properly assess chronic O₃ stress effects on tree leaf litter quality. Liu *et al.* (2005) reported increased condensed tannin concentrations under O₃ exposure (Table 3) and interestingly, this stimulating effect of O₃ was most apparent in the combined CO₂+O₃ treatment. However, other studies show that O₃ does not significantly affect the condensed tannins (Lindroth *et al.* 2001; Table 3), and under the combination treatment the CO₂ effect seems to dominate over the O₃ effect (Parsons *et al.* 2004; Kasurinen *et al.* 2007). Of other C-based compounds, high O₃ levels (100 ppb) seem to induce stress lignin production in foliage (Cabané *et al.* 2004), but when O₃ stress is milder, the O₃ response of lignin (Table 3) and other cell wall components has ranged from no change to slight decrease or increase, the first response pattern being the most commonly observed one (Boerner and Rebbeck 1995; Kim *et al.* 1998; Table 3). As with total phenolics, under the combination treatment the O₃ effect is usually dominated over by the CO₂ effect and therefore the combination treatment effects on cell wall chemistry are largely similar as the effect of elevated CO₂ alone (Parsons *et al.* 2004; Kasurinen *et al.* 2006). Limited data also indicates that monoterpene concentrations do not change due to O₃ stress either under ambient or elevated CO₂ levels (Kainulainen *et al.* 2003).

Decomposition and soil food web responses

A few studies indicate significant O₃-induced decreases in the subsequent decomposition rates in ambient O₃ environment (Findlay and Jones 1990; Findlay *et al.* 1996), but other studies report no change (Boerner and Rebbeck 1995; Scherzer *et al.* 1998; Table 4) or genotype-dependent slow down in the decomposition rates, but only after several exposure seasons (Table 4). Interestingly, Parsons *et al.* (2004) did not observe any changes in the absolute mass loss of O₃ exposed leaf litter when paper birch litter was incubated in its original plot (elevated O₃ plot) or common garden (ambient air) or when leaf litter from control trees were incubated in high O₃ plots (Table 4). However, when leaf litter from control trees was incubated in high O₃+ambient CO₂ plots, the relative mass loss rates (k-values) were significantly accelerated leading to shortened maximum residence times (time-to-95%-loss) (Parsons *et al.* 2004). The effects of combination treatment on the absolute mass

Table 4 Summary of O₃ effects on subsequent leaf litter decomposition (absolute mass loss). Symbols: ↔ = no change, ↓ = decrease, ** = genotype-dependent O₃ response. Only data from OTC- and FACE-studies with soil-growing trees and naturally abscised leaf litter was included. Abbreviation nd = not determined.

Tree species/community	Mass loss		Reference
	Decomposition environment		
	Ambient O ₃	Elevated O ₃	
<i>Betula papyrifera</i> (Native placement + Common garden)	↔	↔	Parsons <i>et al.</i> 2004
<i>B. papyrifera</i> (Common substrate)	↔	↔	Parsons <i>et al.</i> 2004
<i>B. pendula</i> (Exp. 1)	↔	nd	Kasurinen <i>et al.</i> 2006
<i>B. pendula</i> (Exp. 2)	↔	nd	Kasurinen <i>et al.</i> 2006
<i>B. pendula</i> (Exp. 3)	↓**	nd	Kasurinen <i>et al.</i> 2006
<i>Pinus sylvestris</i>	↔	nd	Kainulainen <i>et al.</i> 2003

loss were similar to that of CO₂ alone meaning that elevated O₃ did not modify the negative CO₂ effects on leaf litter decomposition in the original or common garden placement, and that the stimulating effect of elevated O₃ environment itself on k-values of common substrate was not manifested under elevated CO₂ (Parsons *et al.* 2004). On the other hand, Kim *et al.* (1998) observed that elevated O₃ concentrations both during the litter production (quality effects) and decomposition (environment effects) slowed down the mass loss rates of broomsedge-blackberry litter mixture. The authors (Kim *et al.* 1998) suggested that the reduced decomposition rates of litter collected from O₃ exposure were related to increased lignin concentrations in blackberry leaves, whereas the possible explanations for the retarded overall decomposition in high O₃ environment could be decreased microbial activity (Larson *et al.* 2002; Phillips *et al.* 2002) or altered fungal community composition (Chung *et al.* 2006) in soil. Furthermore, FACE studies show that under the combination exposure elevated O₃ can suppress the CO₂-induced enhancement of soil microbial activity and thereby C compound decomposition (Larson *et al.* 2002; Phillips *et al.* 2002).

Soil fauna responses to elevated O₃ are less studied and more variable. Kasurinen *et al.* (2007) observed that leaf litter exposed to elevated O₃ alone and in combination with CO₂ did not consistently alter woodlice leaf litter consumption rates in a laboratory study. In contrast, Loranger *et al.* (2004) have demonstrated that elevated O₃ exposure as an individual treatment can decrease the abundance of microbial-feeding mites (Acari) in the experimental plots, but that under combination treatment both O₃ and CO₂ effects are negated and result is no net change in the total soil fauna abundance.

Boerner and Rebbeck (1995) reported that in their OTC experiment more N was mineralized from leaf litter of yellow-poplar produced under ambient and doubled O₃ air than from that produced under O₃-filtered air, but there was no significant difference between the ambient and doubled O₃ treatments. In a study with single and combination treatments, Parsons *et al.* (2004) did not find any clear O₃ effect on N dynamics in decomposing leaf litter while in the combination treatment elevated CO₂ effect dominated over the O₃ effect so that N dynamics were similar to that in single exposure of CO₂. Kainulainen *et al.* (2003) and Parsons *et al.* (2004) did not either find any significant O₃ or CO₂+O₃ treatment effects on the C-based secondary compounds during their field litter incubations. On the other hand, in a FACE study with three temperate deciduous species (trembling aspen, paper birch and sugar maple), elevated O₃ has been observed to decrease gross N mineralization rates (Holmes *et al.* 2003), especially in combination with elevated CO₂ (Holmes *et al.* 2006). Although the above result is not universally applicable, it suggests that O₃ may have a negative feedback on N cycling in some temperate forest ecosystems and this feedback may be enhanced by the simultaneous increase in atmospheric CO₂.

CONCLUSIONS

Elevated CO₂ increases leaf litter quantity entering the soil, and at the moment FACE studies indicate that this CO₂ stimulation of litter production may be maintained over several years through increased N uptake or NUE. However, if N redistribution in soils occurs (i.e. plant available N decreases), positive effects of elevated CO₂ on forest NPP and litter production may be declined over time. Elevated O₃ alone can decrease leaf litter quantity and increase leaf abscission rates, but due to a paucity of data an average trend in the combined exposure cannot be stated yet. However, a majority of the current CO₂ and O₃ litter studies have been conducted with young trees. Since tree responsiveness (e.g. growth responses) to CO₂ and O₃ may be partially dependent on tree developmental stage or age, more litter and NPP studies with mature trees or non-expanding forests are still needed.

Both elevated CO₂ and O₃ have the potential to alter leaf litter quality, especially in deciduous trees. However, the effects of CO₂ and O₃ on litter chemistry are inherently more difficult to detect than those on green foliage mainly because both gases can also affect leaf senescence processes and thereby increase variability. Elevated CO₂ can especially change the C:N-ratio as both the N and phenolic compound concentrations are usually altered (concentrations of N decreases and e.g. that of condensed tannins increases), whereas N resorption efficiency is not affected by the CO₂ enrichment. Based on recent studies, elevated O₃ effects are mainly seen in litter nutrient concentrations (e.g. P concentrations and some micronutrient concentrations may decrease), whereas O₃ stress effects on leaf litter non-structural and structural C compounds are less apparent or became only apparent, when O₃ concentration is higher than 100 ppb. Under the combined exposure, the CO₂ effect usually dominates over the O₃ effect, and leaf litter quality is usually similar although not necessarily identical to that observed in elevated CO₂ alone.

A majority of the field incubation studies have not found any significant CO₂- or O₃-induced reductions in the subsequent decomposition rates (i.e. mass loss, C and N dynamics). The lack of responses in litter decomposition studies is probably partly due to the low sensitivity of the techniques used such as litter bag studies. However, there are some exceptions, as recent longer-term exposure studies with soil-growing deciduous trees have revealed that both elevated CO₂ and O₃ effects on the subsequent decomposition rates can be negative or that high CO₂ or O₃ environment itself can also modify decomposition rates. Some CO₂ effects on initial leaf litter quality (e.g. decreased N concentrations, increased condensed tannin concentrations) may persist throughout the decomposition, whereas elevated O₃ effects on N and C dynamics in the decomposing leaf litter have been negligible. Under combined exposure no general decomposition response pattern has yet emerged although some studies indicate that under the combination treatment the N dynamics seems to be similar as that in elevated CO₂ alone.

Interestingly, recent FACE experiments also show that especially the functioning of primary decomposers (mic-

robes) may be affected by the increasing CO₂ and O₃ levels. The few available studies show that elevated CO₂ can stimulate microbial activity, whereas O₃ dampens it, and under combined exposure, O₃ can suppress the CO₂-induced enhancement of soil microbial activity. However, recent FACE studies do not show consistent evidence of microbe-mediated progressive nitrogen limitation in temperate forests due to CO₂ increment. There is also a paucity of data regarding the O₃ effects alone and in combination with CO₂ on soil N cycling especially in boreal forests.

Taken together, the experiments represented here show that there are some combined effects that cannot be predicted on the basis of single exposures and therefore, in order to study realistically climate change effects on decomposition and nutrient cycling processes in the forests, effects of increasing CO₂ and O₃ must be studied in combination. In addition, in the future the ongoing global climatic warming may modify the CO₂ and O₃ responses in forests. For instance, temperature rise can increase soil drought and thereby either enhance or control over the effects of CO₂ and O₃ on decomposition and nutrient cycling processes.

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