

Regulatory Role of Mineral Elements in the Metabolism of Medicinal Plants

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ABSTRACT

Minerals have a diversified role in medicinal plant metabolism. Severity or scarcity of these causes multifarious effects in plant metabolism. Each and every aspect of plant biochemistry, physiology, anatomy, etc. is affected due to mineral nutrient composition of soils. Medicinal plants inherit resistance due to biosynthesis of bioactive substances (secondary metabolites) against the various types of diseases caused due to fungus, bacteria, viruses, mycoplasmas, insects and pests. The concentration of these minerals of both group i.e. activators or inhibitors present in the soil play a vital role in secondary plant metabolism. Minerals also play a major role in the reproduction of these medicinally important plants. Bioactive molecules of medical relevancy such as alkaloids, flavonoids, lignans, lipids, carbohydrates, resins, glycosides, phenolic compounds, volatile oils, vitamins, tannins etc. produced through various biosynthetic pathways of plants are a boon to urban, hilly and remote population of each nation. However, soils with different compositions of mineral elements adversely influence the metabolic activities of such valuable medicinal plants. Various physiological activities are governed through important mineral elements present in soils from where these are transferred to area where their need arises. So, accumulation and biosynthesis of these bioactive molecules in a plant system are widely dependent on the availability of mineral elements in the soil. Different developmental stages of the medicinal plants need supplementation of different macro- and micro- elements during its various growth and biosynthesis steps. This review deals with regulatory role of various mineral elements in each biological activities of different medicinal plants viz. *Papaver*, *Catharanthus*, *Withania*, etc.

Keywords: alkaloids, bioactive molecules, growth, nutrition, pathways

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INTRODUCTION

Minerals have a profound effect on the metabolic activities in medicinal and aromatic plants (MAPs) that represent a consistent part of the natural biodiversity endowment of

many nations of the world (Okigbo *et al.* 2008). The role and contributions of various mineral elements can be revealed through their regulatory role played in metabolism of medicinal and aromatic plants. Innumerable substances of immense importance are synthesized such as sugars,

starch, cellulose, acids, lignin, tannins, amino acids, proteins, amides, etc during metabolic processes in plants (Soetan *et al.* 2010). Most of the medicinal plants produce secondary metabolites as for instance morphine in *Papaver*, withanolides in *Withania*, vincristine and vinblastine in *Catharanthus*, etc. which are a boon to human health. The utilization of MAPs as a source of fuel, building material, food, fodder and fibre, in every country has, however, led to a reappearance of natural product-based industries and pharmaceutical products. This had been spurred by the interests of the developed countries for traditional medicine and natural products (Okigbo *et al.* 2008). Today, there is an increasing requirement for plant-based medicines, health products, pharmaceuticals, food supplements, cosmetics, etc. in the international market. The international market of medicinal plants is over 60 billion US\$ per year, which is growing at the rate of 7% per annum. The present export of herbal raw materials and medicines from India is about 100-114 million US\$ per year approximately. India is one of the major exporter of crude drugs among six developed countries viz. USA, Germany, France, Switzerland, U.K. and Japan, and shares 75-80% of the total export market between them (Chatterjee 2001). Global market size for herbal and medicinal plants is projected to reach US\$ 5 trillion by 2050 (www.smeworld.org 2010).

During plant growth, there are continuous processes of building up of complex compounds of carbon (C) and nitrogen (N) and their breakdown into simple substances, in which water (H_2O) and oxygen (O) are intimately concerned. The chief processes involved in plant metabolism viz. absorption, C assimilation, photosynthesis, formation of protoplast, transpiration, respiration, translocation, storage, etc. are regulated by micro- and macro-elements readily available in different forms in the soil (Soetan *et al.* 2010). Supplementing nutrient requirement of medicinal crops through organic manures plays a key role in sustaining soil fertility and crop productivity (Patra *et al.* 2000). With growth cycles, there are certain well defined chemical cycles of nutrient elements and their elaborated products in the leaves, stems and roots, etc. Even the presence of adequate quantities of plant nutrients in the soil does not guarantee its absorption by the plant roots. For example, the latter condition is found in poorly aerated soils, where lack of O₂ near the roots may prevent them from being actively absorbed by the roots (Welch and Graham 2004; Soetan *et al.* 2010).

It is well known that certain elements are necessary for the healthy growth and proper functioning of plant metabolism (Ozcan 2003). They are sometimes referred to as essential elements; some of them are needed relatively in larger quantities and while others in very small amounts. Thus, the former elements are referred to "major" elements and the latter ones as "minor" or "trace" elements or as "micro-elements" (Eruvbetine 2003). The terms "major" and "minor" do not refer to the relative importance of the functions of elements in plant growth, and for this reason the term "trace" element is preferable for the latter class. In addition, there are certain other elements, such as sodium (Na), chlorine (Cl) and silicon (Si), which produce beneficial effects on the growth of certain plants but have not so far been shown to be absolutely essential to growth. The element aluminium (Al) is of general occurrence in plants, but it seems to be without direct nutritional value, although aluminum sulfate is used, because of its acidifying properties, to change the color of hydrangeas growing on alkaline soils from pink to blue. Al may exert indirect influences on nutritional processes as well (Mossor-Pietraszewska 2001). Other elements often occur in plants but they are not known to serve any useful function and are frequently referred to as poisons or toxins to the plant. The nutrient elements can only be absorbed by plants when present in certain definite forms, as for example N in nitrates and ammonium salts, phosphorus (P) in phosphates, calcium (Ca), magnesium (Mg) and potassium (K) in their salts as sulfates or chlorides, sulphur (S) in sulphates, iron (Fe) in ferrous or ferric

salts (more readily from ferrous salts), manganese (Mn) in magnanimous salts, boron (B) in borates, copper (Cu) and zinc (Zn) in their salts, and molybdenum (Mo) in molybdates (Soetan *et al.* 2010; Mazid *et al.* 2011).

Among the various macro-elements required for plant metabolism, N is a major constituent of the most important substances found in plants. N constitutes 40 to 50% of the dry matter of protoplasm, proteins, chlorophyll, amino acids, amides and alkaloids (Oaks 1994; Lea 1997; Fuentes *et al.* 2001). P is closely concerned with the vital growth processes in plants. It is a major constituent of nucleic acid and nuclei (Soetan *et al.* 2010). In seeds, it is related to metabolism of fats. It is also required in root development and ripening of seeds and fruits. Ca and Mg are the constituents of cell wall and chlorophyll; and act as carrier of P element (Belakbir *et al.* 1998; Olaiya 2006). K is required in the formation of carbohydrates and proteins. It is used in regulation of H₂O relations within the plant cell and H₂O loss by plants through transpiration. Besides, it is a catalyst and condensing agent of complex substances and an accelerator of enzyme action. S is a constituent of proteins and is also connected with chlorophyll formation (Hell 1997; Soetan *et al.* 2010; Waraich *et al.* 2011).

Micro-elements, required for plant metabolism, include Fe, B, Zn, Cu, Si, nickel (Ni), cobalt (Co) and Cl. Fe and Mn are used for chlorophyll formation (Rouault 2005); B is used as a catalyst or reaction regulator (Bolaos *et al.* 2000; Olivares *et al.* 2011); Zn and Cu are used as catalysts and regulators (Aziz *et al.* 2010); Si is used in deposition of cell walls (Sacala 2009); Mo is used as a cofactor to enzymes important in building of amino acids (Kaiser *et al.* 2005); Ni is used for activation of urease in higher plants and activates several enzymes involved in a plant metabolism. It is also a substitute element for Zn and Fe, which act as cofactors in several enzymes in the lower plants. Co has proven beneficial in some plants such as legumes, where it is required for N fixation (Aziz *et al.* 2007). Vanadium (V) is required by some plants at very low concentrations. Na is used as replacer of K for regulation of stomatal opening and closing (Pardo and Quintero 2002). The evidence of the role of Cl in plants is somewhat contradictory and no general statement can be made (Runguphan *et al.* 2010). In tobacco, it has been shown to increase H₂O content of the tissues and affect carbohydrate metabolism, leading to an accumulation of starch in the leaves. The element remains present in plants as chloride and is totally soluble. Toxic effects of metals in plants may be produced both by essential and non-essential elements. The major nutrients are less toxic than the trace elements. Indeed, for the major nutrients, there exists a fair safety margin for excess or "luxury" consumption, but for the trace elements the margin is very narrow. Similar conditions exist in relation to non-essential elements; some plants tolerate fairly large amounts of elements such as Na or Cl, but are injured by relatively small amounts of elements like arsenic (As) or chromium (Cr) (Epstein and Bloom 2004). Thus, the present review explains regulatory role of each essential and non-essential mineral element in the common metabolic activities like photosynthesis, respiration, ion uptake, protein synthesis, alkaloid biosynthesis etc., occurring in medicinal plants with the emphasis given to selected medicinal plants such as *Papaver somniferum*, *Catharanthus roseus*, *Withania somnifera*, *Nicotiana*, etc.

REGULATORY ROLE OF MINERAL ELEMENTS IN PHOTOSYNTHESIS AND RESPIRATION WITH REGARDS TO MEDICINAL AND AROMATIC PLANTS

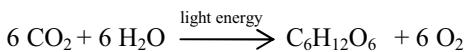
One of the most fundamental and least understood areas of plant physiology and biochemistry is the relationship between quantities of essential mineral nutrient elements and metabolic changes occurring in plants. A living plant is a complex biological system composed of several chemical constituents. C, hydrogen (H), O, S and N are the major

constituents of organic material, which are also involved in enzymatic processes. C and O have a significant role as components of carboxylic group. H and O are useful in oxidation-reduction processes and N is used in biological nitrogen fixation (Salisbury and Ross 2002).

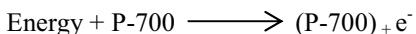
Role of macro-elements

1. Carbon, hydrogen and oxygen

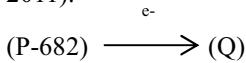
The assimilation of C in form of carbon dioxide (CO_2) takes place in plants and the process is called carboxylation, which is a basic mechanism by which CO_2 is fixed in photosynthesis. The first CO_2 acceptor is RuBP (Ribulose bisphosphate), a three C compound in C₃ plants. This is one of the important reactions of Calvin Cycle (Soetan *et al.* 2010; Murchie and Niyogi 2011). In C₄ plants, CO_2 acceptor is phosphoenolpyruvate (PEP), a four C compound (Edwards *et al.* 2010; Gowik and Westhoff 2011). The reverse process by which CO_2 is liberated is known as decarboxylation; for example, the release of CO_2 from malic acid to form pyruvic acid is due to a decarboxylation reaction. The conversion of light energy into chemical energy is closely related to the conversion of CO_2 into organic compounds (Cruz *et al.* 2005).



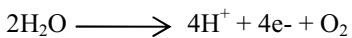
In modern terminology, the process of absorbing electromagnetic radiations by the pigment system and its conversion into chemical energy which can be made available for growth in a particular environment is termed as photosynthesis (Pottosin *et al.* 2005). In higher plants, light absorption is brought about by chlorophyll and carotenoids. Photosynthesis begins with the absorption of light by these pigments, which induces an electron flow that results in conversion of light energy into chemical energy. In higher plants, two photosystems are responsible for such energy conversion; photosystem I and II (Cruz *et al.* 2005). Each of these photosynthetic units consists of about 400 chlorophyll molecules along with additional pigments such as carotenoids. In photosystem I (PS-I) a special chlorophyll molecule is involved known as pigment 700 (P-700) (Yokthongwattana and Melis 2006; Murchie and Niyogi 2011).



This is the basic process that initiates electron flow. The electron ejected from P-700, thus, moves against an electrical gradient. Photosystem II (PS-II) functions in an analogous way to PS-I. In PS-II, the electron emitter is a chlorophyll molecule P-682 (Cruz *et al.* 2005; Kramer and Evans 2011).



These two photosystems function in series and constitute the components of electron transport pathway, transferring the electrons from H_2O to (Nicotinamide adenine dinucleotide phosphate) NADP^+ . Thus, H_2O being the ultimate electron donor and NADP^+ the electron acceptor in the overall process. PS-II is closely associated with splitting of H_2O molecule, which serves as electron donor (Shikanai 2007).



Photolysis of H_2O i.e. splitting of H_2O molecule makes use of Mg (Mg^{2+}) and Cl (Cl^-) ions. The whole process does not seem to be legitimate to describe here in details. Plastocyanin (a Cu containing acidic protein) and ferredoxin (Fe-S protein) plays critical role in this electron transport system (Diner and Babcock 1996; Aubry *et al.* 2011). In photosynthetic energy conversion, light induces an electron flow that provides sufficient energy for the synthesis of

adenosine triphosphate (ATP) and reduction of NADP^+ . In the whole process, another Cu containing compound plastocyanin and an Fe containing compound cytochrome play vital roles (Sacksteder *et al.* 2000).

The next major metabolic process being the cellular respiration (oxidative metabolism) which is a set of metabolic processes and reactions are executed within the cell of an organism in order to convert biochemical energy, derived from the nutrients, to ATP (Soetan *et al.* 2010). The ATP is a major source of energy for cellular reactions taking place in all form of life. There are two types of cellular respiration: aerobic and anaerobic cellular respiration. Both animals as well as plants, execute cellular respiration to produce energy. They use nutrients such as glucose, amino acids and fatty acids to produce energy. The common oxidizing agent in this process is molecular O. Glycolysis being the first step of cellular respiration, is the process whereby the glucose is chemically modified to pyruvate. This process occurs in cytosol of the cell and is carried out in the absence of O. During this process, energy is released from glucose compounds in the form of 2 NADH molecules and 2 ATP molecules. Pyruvate decarboxylation is the second stage of cellular respiration. In this process, pyruvate is decarboxylated and added to coenzyme A (CoA) in order to form acetyl CoA. It is an important stage in the cellular respiration process, which forms a link between metabolic pathways of glycolysis and the Krebs's cycle.

The Krebs's cycle comprises a series of steps, which oxidize acetyl CoA molecule. Unlike glycolysis, the Krebs's cycle requires O for its functioning and is catalyzed by various series of enzymes. Two complete turns of Krebs's cycle produces 4 CO_2 molecules, 2 ATP molecules, 6 NADH₂ and 2 Flavine adenine dinucleotide (FADH₂) molecules (Soetan *et al.* 2010). The final stage of cellular respiration is electron transport chain (ETC), which produces the remaining 32-34 ATPs. The ETC comprises of electron-carrying proteins, present on the internal membrane of mitochondria. These proteins transfer electrons from one self to another. These electrons are finally added to O, which is reduced to form H_2O . In this process, ATPs are produced by pmf, a source of potential energy created by the gradient that is formed when protons move across the biological membrane. Simply the ETC triggers a pH gradient through which ATP is produced by the process known as chemiosmosis.

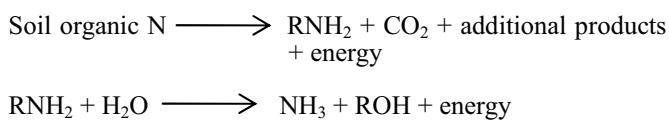
A short-term exposure to CO_2 enhanced the rate of photosynthesis and *vice versa*, while an intermediary exposure of CO_2 had variable effect on the rate of photosynthesis in C₃ plants (Makino and Mae 1999). The suppression of photosynthesis by CO_2 enrichment is always associated with decreases in leaf N and Rubisco contents. These decreases are not due to dilution of N caused by a relative increase in the plant biomass, but occur as a result of decrease in N allocation to leaves at the level of whole plant, and the decrease in Rubisco content is not selective. Accumulation of carbohydrates in the leaves may lead to the repression of photosynthetic gene expression. Besides, the excess starch hinders the CO_2 diffusion through the leaves. In *Papaver setigerum*, increasing CO_2 concentration (300, 400, 500, 600 $\mu\text{mol mol}^{-1}$) had positive effect on various morphological traits such as number of capsules, capsule weight and latex-yield, with an increase of 3.6, 3.0 and 3.7 times, respectively on per plant basis. The secondary metabolic products also responded positively to CO_2 enrichment. The concentration of alkaloids morphine, codeine, narcotine and papaverine also increased in response to enhancement of atmospheric CO_2 . The concentration of major alkaloid i.e. morphine was significantly increased by 10.4, 11.7, 12.9 and 12.4%, respectively for each dose (300, 400, 500 and 600 $\mu\text{mol mol}^{-1}$) of CO_2 (Ziska *et al.* 2008). Similarly, there was a positive effect of increasing atmospheric concentration of CO_2 (950-1050 $\mu\text{mol mol}^{-1}$) on hypericin and pseudo-hypericin content (14-fold increase) in *Hypericum perforatum* (Zobayed and Saxena 2004). There was an increase of 15% in digoxin yield per unit dry weight of plant and 75% increase in plant dry weight production per unit land

area in *Digitalis lanata* by near-tripling of the air's CO₂ concentration (Stuhlfauth and Fock 1990). Similarly, the concentration of atropine (11.9, 14.7, 18.9 mg) and scopolamine (12.9, 20.9, 25.4 mg, respectively) increased in *Datura stramonium* by varying CO₂ concentrations (294, 378 and 690 µmol mol⁻¹) (Ziska *et al.* 2005).

2. Nitrogen, phosphorus and potassium

N is a main constituent of all the amino acids, proteins and coenzymes (Mazid *et al.* 2011; Murchie and Niyogi 2011). P occurs as phosphate or orthophosphate and to a minor extent as pyrophosphate. The process by which plants convert inorganic N and S into organic forms is important for all the animals that depend on plants and microorganisms for their dietary source of organic N and S. N is one of the most widely distributed elements in nature that circulates in atmosphere, lithosphere and hydrosphere. Many factors are involved in the N-turnover, some of which are physicochemical and others biological. As such, dry plant material contains approximately 2 to 4% N. N is an indispensable elementary constituent of numerous organic compounds of general importance (amino acid, protein, nucleic acid, etc.). Both, NO₃⁻ and NH₄⁺ forms of N can be taken up readily by plants from the soil and are metabolized thereafter in the plant cells. The rate of uptake of NO₃⁻ is generally very high as plants require large amount of N for their growth and development. It is now established that in this uptake process there are involved both influx and efflux components (Deane-Drummond and Glass 1983). Influx of NO₃⁻ is an active process that involves the movement of NO₃⁻ against the electrochemical gradient. The influx of NO₃⁻ is also dependent on the concentration of NO₃⁻ in the external medium (Lillo *et al.* 2004; Halkier and Gershenson 2006; Soetan *et al.* 2010).

N fixation is the most important process, in which the molecular N of the atmosphere from its inorganic form is fixed and converted into an organic form. Only prokaryotes are capable of assimilating molecular N. In addition, proteolysis, ammonification, nitrification and denitrification are some other processes, which are controlled by microbial activity (Fuentes *et al.* 2001; Stitt *et al.* 2002).



In opium poppy (*Papaver somniferum* L.) suitable form, supply and date of application of N fertilizers play a decisive role in proper growth of poppy plants and its yield. The need of optimal supply of N for poppy plants begins shortly after seed germination and lasts till the stage of formation of the generative organs (Yadav *et al.* 1984; Pavlíková and Tlustoš 1994). Yadav *et al.* (1984) suggested that slow release of N had a positive effect on the seed yield of opium poppy. Increase in the seed yield was due to enhancement in the process of photosynthesis, which was accelerated by the addition of N. In *Catharanthus roseus*, the growth of seedlings was improved by increasing N concentration in the pre-transplant fertilizer from 8 to 32 mM, when a peat-based growing medium was used in the experiment (Van Iersel *et al.* 1998). N supply in *C. roseus* also stimulated K and P uptake and their translocation to the leaves (Lata and Sadowska 1996). N is a major limiting nutrient for plant growth; it is assimilated as NH₄⁺ by the converted action of glutamine synthase (GS₁ and GS₂) and glutamate synthase (GOGAT). In tobacco (*Nicotiana tabacum*), Fuentes *et al.* (2001) over expressed GS₁ at the level of RNA and protein which increased the activity of GS upto six times in comparison to control plants which resulted in 70% higher shoot, 100% greater root dry weight and 50% more leaf area than controls. They further concluded that manipulation in GS activity has the potential to maintain crop productivity while reducing nitrogen fertilization and the concomitant

pollution. Omer (1998) and Omer *et al.* (2008) noticed that nitrogen fertilizer was effective in increasing essential oil of *Origanum syriacum* and *Ocimum americanum*, respectively.

The organic forms of phosphate are the compounds in which, orthophosphate is esterified with hydroxyl group of sugars and alcohols or it is bound by a pyrophosphate bond to other phosphate group. A typical example of a phosphate ester is fructose-6-phosphate which is an intermediary metabolite of glycolysis. Such organic phosphates are usually intermediary compounds of metabolism. Phosphate is also bound to lipophilic compounds particularly in phosphatidyl derivatives e.g. lecithin. Nagano and Ashihara (1993) revealed that the metabolic adaptation of inorganic phosphate (P_i) in respiratory pathways in plants was restricted to a limited level and that there were significant effects of long-term P_i starvation on the activities of various enzymes related to respiratory metabolism. They found that only PEP-hydrolyzing enzyme and PEP carboxylase were in higher amounts in P_i-starved cells of *C. roseus*. Activities of other enzymes in the P_i-starved cells were lower than or similar to those of the control cells. Time-course studies indicated that PEP-hydrolyzing activity was inducible by starvation of P_i. However, fluctuations in the activity of PP_i:fructose-6-phosphate i-phosphotransferase during starvation of P_i in levels was similar to those of phosphofructokinase and 6-phosphogluconate dehydrogenase (Williamson *et al.* 2001). In *Plantago ovata*, increasing concentration of P plays a critical role in elevating total yield of the plant which was ascribed to the regulatory role of P in metabolism (Tomar *et al.* 2010). Similarly, P deficiency prevented nodulation or stopped nodule growth and also decreased photosynthesis in *Trifolium repens* (Almeida *et al.* 2000; Asatsuma *et al.* 2003).

The most important compound in which phosphate groups are linked by pyrophosphate bond is ATP. The pyrophosphate bond is an energy rich bond, which on hydrolysis releases 30 KJ Mol⁻¹ energy. The energy produced or released in various metabolic processes is utilized in the synthesis of pyrophosphate bond of ATP. In the form of ATP, the energy can be conveyed to various endergonic processes such as active ion uptake and synthesis of various organic compounds. In the whole process, there is usually an initial phosphorylation reaction which involves transfer of phosphoryl group from ATP to another compound. Another organic compound containing P is phytin which occurs mainly in seeds. Phytin is synthesized during seed formation in plants and occurs in the form of Ca and Mg salts of phytic acid in the seeds. Phytin is regarded as P reserve in the seeds. Phytic acid is the hexaphosphoric ester of inositol (Lott *et al.* 2000; Angel *et al.* 2002; Asatsuma *et al.* 2003; Pelig-Ba 2009).

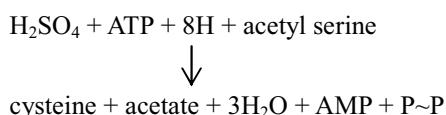
The role of K in increasing photosynthesis of *Medicago sativa* plant can differently be understood as K⁺ did not directly influence PS I or II; instead, it promoted the *de novo* synthesis of enzyme ribulose bisphosphate carboxylase (Peoples and Koch 1979; Demmig and Gummel 1983; Salim 2002). K decreased the diffusive resistance for CO₂ in mesophyll cells. The movement of K⁺ from the thylakoid spaces into the stroma of chloroplasts favors electron flow in the transport chain (Subbarao *et al.* 2003; Horie *et al.* 2007; Haro *et al.* 2010). Depolarization of the plasma membrane by K⁺ uptake has a direct influence on phloem loading. Numerous researchers have shown that K⁺ enhances the translocation of photosynthates. K not only promotes translocation of newly synthesized photosynthates but also has a beneficial effect on the mobilization of stored material. Flores *et al.* (2000) emphasized that a decrease in K content of *Catharanthus* due to sodium chloride (NaCl) treatments may be due to the toxic effect of NaCl on plant growth or due to competition by other ions, which, in turn, exercised a regulatory control on K uptake.

The decomposition of plant and animal materials leads to the production of organic compounds i.e. humic substances. To ameliorate soil acidity and improve soil structural stability, humic acid and their salts derived from coal and

other sources may provide a viable alternative to liming. Zaghloul *et al.* (2009) reported that the application of potassium humate (0.0, 0.5, 1.0, 1.5, 2.0 and 2.5% K-humate) leads to increase oil content in *Thuya orientalis*. Said-Al *et al.* (2009) concluded that foliar application of K-humate promoted growth and possessed the best oil percentage in oregano plant. The increase in permeability of plant membranes due to humate application results in improved growth of various groups of beneficial microorganisms, accelerate cell division, increased root growth and all plant organs for a number of horticultural crops and turf grasses, as well as, the growth of some trees (Russo and Berlyn 1990; Sanders *et al.* 1990; Poincelot 1993).

3. Sulphur, calcium and magnesium

S occurs in the soil in inorganic and organic forms. In most soils, organically bound S provides major S reservoir (Reisenauer *et al.* 1973; Scott and Anderson 1976). Soil organic S can be divided into 2 fractions: C bonded S, and non-C bonded S. The latter fraction is made up of phenolic and choline sulphates and lipids (Freney and Stevenson 1966). The inorganic form of S in soil consists mainly of SO_4^{2-} , which is mainly absorbed by plants. S taken up by plant cells, must be reduced as in majority of S containing molecules S is present in reduced form (Saito 2000; Mazid *et al.* 2011). The S containing organic compounds include the amino acids cysteine, cystine and methionine as well as the proteins containing these amino acids. The first step in S assimilation is the incorporation of S between H_2SO_4 (sulphuric acid) and ATP. The sulphuryl group of H_2SO_4 replaces the pyrophosphoryl group of ATP, thus forming adenosine phosphosulphate and pyrophosphate, which is catalysed by an enzyme ATP sulphurylase. The sulphuryl group of adenosine phosphosulphate (APS) is transferred to an SH-carrier complex which is then reduced to -SH group, the reducing power provided by the ferredoxin. In a further step, the -SH group of the carrier complex is transferred to acetyl serine leading to regeneration of original carrier SH-complex. Acetyl serine then split into cysteine and acetate. This splitting step requires two additional reducing equivalents, which also probably originate from ferredoxin (Saito 2000).



This equation shows that sulphate reaction needs energy in the form of ATP and reducing equivalents. Hence, the reduction process depends on photosynthesis and especially on the level of ATP (Schiff and Hodson 1973). Sulphate is mainly reduced during the light period because sulphate reducing enzymes are located in the chloroplast membrane. Whether other organelles are also capable of reducing sulphate, is not known. Modern agriculture requires adequate fertilization of S to achieve maximum crop yield and performances (Crawford *et al.* 2000). S^{2-} a major form of SO_4^{2-} in aqueous phase of apoplast, may reduce photosynthesis rate and thereby crop yield through inducing reactive oxygen species (ROS), involved in S^{2-} -induced stress, and the S^{2-} -induced enhancements in levels of ROS (Li *et al.* 2007).

The role of Ca in membrane stability is by far the most important process but other metabolic processes are also dependent on Ca availability (Rengel 1992; Marschner 1995; Tuna *et al.* 2007). The feature includes a breakdown in the compartmentation of cell and an increase in cellular respiration, followed by the leakage of endogenous respiratory substrates from the vacuole to the respiratory enzymes in the cytoplasm (Bangerth *et al.* 1972). Most of the Ca present in plant tissues is located in the apoplast and vacuoles. The Ca concentration of cytoplasm remains low, ranging from 10^{-6} to 10^{-8} M (Marme 1983). The maintenance of this

low cytosolic concentration of Ca is of vital importance for the plant cells (Hanson 1984), for there is now evidence that Ca may inhibit the activity of various enzymes located in the cytoplasm (Gavalas and Manetas 1980). The same is also true for the chloroplast enzymes. The low Ca concentration not only prevents inhibition of enzyme activity, but it also inhibits precipitation of P_i as Ca-phosphate and the competition of Ca^{2+} with Mg^{2+} for binding sites of the enzymes (Hepler and Wayne 1985). The maintenance of low Ca concentration is achieved by the mechanisms which pump Ca^{2+} out of the cytoplasm into the apoplast or into the vacuole. Some of the Ca^{2+} is also sequestered by the endoplasmic reticulum and mitochondria, organelles relatively rich in Ca^{2+} .

Ca concentrations in the mitochondria are much higher than in the cytoplasm. Mitochondrial enzymes may be directly activated by Ca^{2+} without the help of calmodulins (Marme 1983). One such enzyme which occurs in the mitochondria is glutamate dehydrogenase, which brings about the deamination of amino acids. Franklin-Tong *et al.* (2002) demonstrated that increase in free cytosolic Ca^{2+} could be triggered by self-incompatibility (SI) response in the pollen of incompatible *Papaver rhoes* (the field poppy). However, one key question that has not been answered is whether extracellular Ca^{2+} may be involved in this regard. To address this question, they used an ion-selective vibrating probe to measure changes in extracellular Ca^{2+} fluxes around pollen tubes of poppy. They confirmed that there was an oscillating Ca^{2+} influx directed at the apex of the pollen tube and also provided the evidence that Ca^{2+} influx occurred at the shanks of pollen tubes. Secondly, upon challenge with self-incompatibility (S) proteins, there was a stimulation of Ca^{2+} influx along the shank of incompatible pollen tubes, approximately 50 mm behind the pollen tube tip. This demonstration of SI-induced Ca^{2+} influx suggested a role for influx of extracellular Ca^{2+} in the SI response.

A major proportion of total plant Mg i.e. often over 70% is present in diffusible form and associated with inorganic anions and organic acid anions such as malate and citrate and also found associated with indiffusible anions including oxalate and pectate (Kirkby and Mengel 1976). The chief importance of Mg is due to its occurrence at the centre of the chlorophyll molecule. The fraction of the total plant Mg associated with chlorophyll, however, is relatively small, ranging 15 to 20% (Neales 1956). Besides its function in chlorophyll molecule, Mg^{2+} is required in other physiological processes. One major role of Mg^{2+} is that it acts as a cofactor in almost all the enzymes activating phosphorylation processes. Mg forms a bridge between pyrophosphate structure of ATP or ADP and the enzyme molecule. The activation of ATPase by Mg^{2+} is brought about by this bridging function (Balke and Hodges 1975). In addition to phosphokinases some dehydrogenases as well as enolase are also activated by Mg^{2+} . In these enzymes, however, Mg reaction is not specific and Mn^{2+} is often a more efficient activator.

Mg^{2+} also plays a vital role in the aggregation of ribosomes. Mg^{2+} concentrations also modulate ionic currents across the chloroplast and vacuolar membranes and might, thus, regulate ion balance in the cell and stomatal opening. The significance of Mg^{2+} in homeostasis has been particularly established with regard to the role of Mg^{2+} in photosynthesis (Panayotov 2004, 2005; Panayotov *et al.* 2005; Kostova *et al.* 2008). Fluctuations in Mg^{2+} level in the chloroplast regulate the activity of key photosynthetic enzymes. Relatively little is known of the proteins mediating Mg^{2+} uptake and transport in plants. The plant vacuole seems to play a key role in Mg^{2+} homeostasis in plant cells. Physiological and molecular evidences indicate that Mg^{2+} entry into the vacuole is mediated by $\text{Mg}^{2+}/\text{H}^+$ exchangers (Shaul 2002; Hattori *et al.* 2007).

A key reaction of Mg^{2+} is the activation of ribulose bisphosphate carboxylase. Light triggers the import of Mg^{2+} into stroma of the chloroplast in exchange of H^+ , thus providing optimum conditions for the carboxylase reaction.

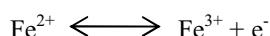
The favorable effect of Mg^{2+} on CO_2 assimilation and related processes, such as sugar and starch production, are probably due to the consequences of this activation of ribulose bisphosphate carboxylase. According to Barber (1982), Mg^{2+} is the most important cation species neutralizing the indiffusible anions of the thylakoid membrane. Skarpa *et al.* (2008) demonstrated the effect of Mg applied in the form of Mg nitrate to the soil supplemented with cadmium (Cd) on yield as well as qualitative and quantitative parameters in *Papaver somniferum* L. During the vegetative period, Mg fertilization had a positive effect on Mg and Ca concentrations in the plant. Mg application also had a synergistic effect on N uptake and increased its content in the plant. In variants where Cd was supplemented, its content in the plants considerably increased. With Mg fertilization, the yield of poppy seeds increased when compared with the unfertilized variants; both in the variant with natural Cd content (3.6%) and the variant that was supplemented with Cd (19.9%). Cd had a synergistic effect, enhancing a better utilization of nutrients and resulting in the higher seed yield. As a result of Mg application the amount of morphine in poppy straw increased both in variants with natural and supplemented Cd content.

Role of micro-nutrients

1. Manganese, iron, copper

In its biochemical function, Mn^{2+} resembles with Mg^{2+} and both form an ATP bridge with the enzyme complex (phosphokinases and phosphotransferases). In the tricarboxylic acid cycle (TCA cycle), two of the functional enzymes i.e. decarboxylases and dehydrogenases are also activated by Mn^{2+} , although it appears that in most cases Mn^{2+} is not specific for these enzymes and can be substituted by Mg^{2+} . For the Mn^{2+} to be effective, it has to be utilized preferentially, as it occurs in the activation of chloroplast RNA polymerase. Activation is brought about by either Mn^{2+} or Mg^{2+} but at low concentrations (1 mM or less); however, the former is much more effective. The peroxidase enzyme activity is enhanced in contrast to catalase enzyme in Mn-deficient tissue (Husted *et al.* 2009). This increase in the activity of peroxidase is probably associated with the high activity of IAA oxidase, which is observed in Mn-deficient plants (Chatterjee *et al.* 1994). Peroxidases are most important constituents of IAA oxidizing system (Jansen *et al.* 2001). The most documented role of Mn in green plants is in the H_2O splitting and O evolution system in photosynthesis (Panayotov 2004, 2005; Panayotov *et al.* 2005). It has now been confirmed that Mn is required both by higher and lower plants, as it appears that a manganese protein complex catalyzes the O₂ evolution (Papadakis *et al.* 2007). This tightly bound Mn complex has not as yet been isolated and its structure is until now unknown. The reaction has been depicted by Edwards and Walker (1983) with a minimum requirement of 4 Mn atoms at each reaction centre of photosystem II. From this scheme, it follows that when Mn is deficient the ETC in the light reaction is seriously disturbed resulting in the occurrence of other reactions including photophosphorylation and the reduction of CO₂ (Apel and Hirt 2004; Moller *et al.* 2007). As a consequence of absence of Mn, nitrite (NO_2^-) and sulphate are detrimentally affected. When NO_2^- reduction is impaired the accumulated NO_2^- can exert a feed back control on NO_3^- reductase activity so that NO_3^- is also accumulated. This is one reason why NO_3^- accumulation is sometimes observed in Mn-deficient plants. Pande *et al.* (2011) investigated the effect of varying Mn concentration (0.0, 2.5, 5.0 and 10.0 mg/kg) on medicinally important plant *Mentha spicata* and concluded that optimal level required was 2.5 mg/kg, beyond which growth parameters showed a gradual and significant decline. Highest stem and leaf yield was noticed which may be attributed to proper root development as indicated by its higher root weight. Mn supply improves the root growth and its activity as reported by Mou *et al.* (2010).

The tendency of Fe to form chelate complexes and its ability to undergo a valency change are the two important characteristic which underlie numerous physiological effects of Fe (Cohen *et al.* 1998).



The most well known function of Fe relates to the activity of the enzymes in which haem functions as prosthetic groups (Rouault 2005). Here, Fe plays a similar role as does Mg in the porphyrin structure of chlorophyll. These haem enzyme systems include catalase, peroxidase, cytochrome oxidase as well as various cytochromes. The detailed function of the cytochrome in electron transport and involvement of cytochrome oxidase in the terminal step of respiration chain does not seem to be legitimate to describe here. Misra *et al.* (2006), while evaluating the Fe-efficient genotypes of the medicinally important plant *Ocimum*, found an oxido-reducible reaction of peroxidase as well as high bands of peroxidase isoenzymes in OSP-6 genotype for the formation of monoterpane essential oil(s) and possibly the major constituents of eugenol through the high production of photosynthates. Pande *et al.* (2011) investigated the effect of varying Fe concentration (0.0, 5.0, 10.0 and 15.0 mg/kg) on medicinally important plant *Mentha spicata* and found that fresh weight, dry weight, oil yield and chlorophyll content increased significantly with increasing Fe concentration. The optimal dose was 10.0 mg/kg for maximum increment.

Plants require Cu in very less quantity for their proper functioning. The Cu content in most plants generally varies between 2-20 ppm in dry plant material. Enzymatically bound Cu participates in redox reactions which are mostly dependent on the valency change.



In this respect, Cu is similar to Fe, although Cu^+ is much less stable than the corresponding Fe^{2+} . In protein complexes, Cu has particularly a high redox potential. Most Cu enzymes/compounds (plastocyanin, superoxide dismutase (SOD) and amine oxidases) react with O₂ and reduce it to H_2O_2 or H₂O. In Cu containing proteins, three different forms of Cu are distinguished (Sandmann and Bogger 1983). Type I is the blue Cu protein (e.g. plastocyanin), which functions without oxidase activity in one electron transfer. Types 2 are non-blue Cu proteins which are peroxide producing oxidases. Type 3 Cu-containing protein (e.g. phenolase) is non-blue and it functions as a two electron acceptor in oxidation process, which catalyses the oxidation of monophenols to ortho- and di-phenols and then further to o-quinones and H₂O. Cu proteins are also present in multi Cu enzymes containing all the above three types of Cu. These enzymes include ascorbic acid oxidase and lactase catalase (Navarro *et al.* 1997).



Approximately 70% of the total Cu in leaf is bound to various organelles/components of the cell, of which more than half is bound to plastocyanin, a component of ETC of photosystem I. The Cu is also a constituent of other photosynthetic enzymes, e.g. phenolase, SOD and fraction 1 protein, which has RuBP carboxylase and RuBp oxygenase activity (Navarro *et al.* 1997). However, a precise role of Cu in photosynthesis is still unclear. The most common of the three types of SOD isoenzymes contain Cu and Zn. This enzyme protein (Cu-Zn SOD) has a molecular weight of about 32,000 da and contains two Zn and two Cu atoms. SOD occurs in all aerobic organisms and is essential for their survival in O environment (Shikanai *et al.* 2003). They protect the organism from the damage of superoxide radicals, which can be formed when a single electron is transferred to O₂. The high proportion of SODs in leaves appears to be localized in chloroplast which indicates a role of

SODs in protecting photosynthetic apparatus but the details about how this is achieved are still lacking (Jackson *et al.* 1978). Cytochrome oxidase, the terminal oxidase in the mitochondrial transport chain, is one of the best studied Cu containing enzymes. This enzyme transfers electrons directly to molecular O₂ which can be inhibited by CN; however, when this reaction occurs, another Cu containing oxidase, the CN- resistant quinol oxidase comes into action. This relatively recently discovered enzyme, called alternative oxidase, thus, provides a second pathway of ubiquinone oxidation in the mitochondria (America *et al.* 1994). Electrons are believed to be transported directly from quinol substrate to molecular O₂, avoiding the cytochrome. Cu influences both carbohydrate and N metabolism. In the vegetative stage of plants, Cu deficiency can lower the content of soluble carbohydrates as might be expected owing to the specific role of Cu in photosynthesis (Gupta *et al.* 2002). In *Withania somnifera*, due to the application of copper sulphate (CuSO₄) to plants, there was observed accumulation of high amount of Cu in concentration manner; it also decreased the plant fresh weight, shoot and root length, chlorophyll content and carotenoid content. Similarly, few phenolic compounds were also increased due to CuSO₄ application (Khatun *et al.* 2008). Santiago *et al.* (2000) analyzed compartmentation of phenolic compounds in mature leaves of *Phyllanthus tenellus* and their induction by CuSO₄. They noticed that spraying plants with CuSO₄ induced punctated lesions formed by groups of necrotic cells which accumulated brownish substances.

2. Silicon, chlorine and sodium

Si is found as a component of cell wall. Plants supplied with soluble Si produce stronger and tougher cell walls, providing a mechanical barrier against piercing and sucking the plant tissue by the insects. Si significantly enhances the tolerance of plant to heat and drought. Foliar sprays of Si have also shown benefits in reduction of populations of aphids in various field crops. Studies have also revealed that Si could be deposited by the plants at the site of infection by fungus to combat the penetration of cell walls by the fungi. Leaf erectness, stem strength and prevention of Fe and Mn toxicity have been improved by the application of Si. Though the Si is not essential for all the plants but can be beneficial for many plants (Epstein 1994; Hodson *et al.* 2005; Ma and Yamaji 2006). Silicic acid like boric acid, however, reacts with o-phenol such as caffeic acid, a precursor in the biosynthesis of lignin, to form mono-, di- or polymeric Si complexes (Sacala 2009). It has also been noted that Si is somehow linked with the transpiration in the plants. Ample evidence are there that Si, when readily available to plants, plays a large role in their growth, mineral nutrition, mechanical strength, and resistance to fungal diseases, herbivory, and adverse chemical conditions of the medium (Epstein 1994; Ma and Yamaji 2006; Epstein 2009; Sacala 2009).

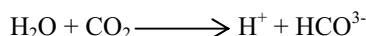
Plant tissues usually contain substantial amounts of Cl, often in the range of 50-500 μmol kg⁻¹ dry weight. These values are comparable to those of macronutrients. Though the demand of Cl for optimal growth is less but its deficiency symptoms occur when Cl content falls below 20 μmol kg⁻¹ dry weight. This clearly established the role of Cl as micronutrient (Clarkson and Hanson 1980). Cl is required in Hill's reaction and the H₂O splitting reaction in photosystem II of photosynthesis (Kelly and Izawa 1978). Cl also enhances both evolution of O₂ and photophosphorylation (Bove *et al.* 1963). In some plants, chloride ion influences photosynthesis indirectly via affecting stomatal regulation in the guard cells. During stomatal opening, inward flux of K⁺ into the guard cells must be accompanied by either accumulation of malate as a counter ion or by the inward flux of Cl. Cl sharply decreased NO₃⁻ uptake by the roots and facilitated the incorporation of N into organic compounds and translocation of N to the leaves in *Catharanthus rheoeus* (Flores *et al.* 2000). However, the studies depicting the role

of chlorine and its compound in plant metabolism are very few, especially in medicinal and aromatic plants.

Na belongs to the group of micro elements and hence, it cannot be expected to have a specific role in the metabolic activities of plants. When Na produces significant effects in plant metabolism, it is often regarded as a replacer of K as it is partly able to replace K if present in excess. Na is ineffective as a substitute for K, even for Na-loving plants such as sugar beet (*Beta vulgaris*), marigold (*Calendula officinalis*) and barley (*Hordeum vulgare*). Na seems to affect the H₂O relations of plants and often enables sugar beet and other crops to withstand drought conditions, which would otherwise produce severe adverse effects. In *Cassia angustifolia*, Arshi *et al.* (2005) found that sodium chloride (NaCl) decreased biomass of root, shoot and leaf significantly, while calcium chloride (CaCl₂) increased the biomass of these plant parts. Combined doses of NaCl and CaCl₂ increased the proline content 8 times, while NaCl alone increased it 5 times. Combined application of NaCl and CaCl₂ decreased the biomass but this effect was lesser than that exhibited by NaCl application alone. K⁺ and Ca⁺ concentration was inhibited with NaCl, while Na⁺ and Cl⁻ concentrations were increased. CaCl₂ and combined treatment (NaCl + CaCl₂) increased the K⁺ and Ca⁺ concentrations as well. Similarly, in some medicinal plants, they also found that NaCl decreased the biomass, length of root and shoot, photosynthetic rate, stomatal conductance, total chlorophyll content, protein content, nitrate reductase activity and reduced N content of leaves, while proline and nitrate content increased (Arshi *et al.* 2002). In *Withania somnifera*, different Na salts viz. NaCl, sodium sulphate (Na₂SO₄) and sodium carbonate (Na₂CO₃) decreased the germination percentage, shoot length, root length, plant dry weight and chlorophyll as well as carotenoids contents of the plants significantly (Jaleel *et al.* 2009).

3. Zinc, molybdenum and boron

Zn facilitates the binding between the enzyme and substrate, bringing about the conformational change in the system. A number of enzymes, including enolase, are activated by Zn²⁺ in more or less the same way they get activated by Mn²⁺ and Mg²⁺. Till now, the only authenticated enzyme that gets specifically activated by Zn²⁺ is carbonic anhydrase (CA). This enzyme catalyzes the following reaction:



It is generally accepted that CA is localized in the cytoplasm but there are also evidences to suggest its presence in the chloroplasts (Findenegg 1979). The enzyme promotes hydrolysis and hydration reactions involving Carbonyl groups (Sandmann and Boger 1983). Jacobson *et al.* (1975) reported its supposed role in mediating a short-term transient pH effect, thus acting as a buffer. Such a role of this enzyme was suggested because of its high concentration in chloroplast stroma. There it protects proteins from denaturation, which can occur as a result of local pH change associated with activation of H⁺ pump and the incorporation of CO₂ into ribulose 1, 5 bisphosphate. The role of CA, however, is still a matter of speculation. Zn is involved in C assimilation, saccharide accumulation, free radical removal, antioxidant enzymes activation, C utilization in terpene biosynthesis and in overall growth of the plants. The requirement of Zn for Japanese mint (*Mentha arvensis*) and its limitations imposed on photosynthetic C metabolism and translocation in relation to essential oil accumulation were shown by Misra and Sharma (1991). Misra *et al.* (2005) showed that in geranium (*Pelargonium graveolens* L.), Zn acts as an antioxidant promoter, apart from its essentiality as micronutrient. Varying concentrations of Zn from 0–1.0 g m⁻³ (Zn₀ to Zn_{1,000}) affected net photosynthetic rate, contents of chlorophyll and essential monoterpene oil(s). Zn_{0.250} resulted in maximum total essential monoterpene oil(s) content i.e. 0.21%. However, Zn_{0.005}–Zn_{0.250} significantly

affected the net photosynthetic rate, contents of chlorophyll and essential monoterpene oil(s). At $Zn_{0.250}$ concentration, maximum peroxidase activity was obtained with the production of biomolecule geraniol. Other enzymes containing Zn include alcohol dehydrogenase, SOD and RNA polymerase (Vallee and Wacker 1970; Sandmann and Boger 1983). Alcohol dehydrogenase catalyzes the reduction of acetaldehyde to ethanol, a reaction in higher plants largely confined to meristematic zones such as root apices. In one of the three types of SODs, the metal prosthetic group includes Zn associated with Cu (Cu-Zn SOD). The metal catalyses the conversion of superoxide radical and O_2^- to hydrogen per oxide and O₂, promoting anaerobic organism to defend themselves from the damage caused by O₂⁻. The function of Zn in this isoenzyme has yet to be established. Foliar fertilization of plants with Zn resulted in an increase of nutrient content in the dry matter of leaves, with the most intensely absorbed element being the Zn itself (Lata and Sadowska 1996).

Mo is an essential element of the respiratory and assimilatory nitrate reductase. The former is confined in denitrifying bacteria and catalyzes the reduction of nitrate to nitrite. Assimilatory nitrate reductase is present in different kinds of organisms such as bacteria, cyanobacteria, algae, fungi and higher plants (Kaiser *et al.* 2005). It is supposed that in oxido-reduction process, Mo facilitates the transport of an electron. Three Mo-enzymes have so far been found in plants (Self *et al.* 2001). Nitrate reductase catalyzes the reduction of nitrate, the key step in inorganic N assimilation. Aldehyde oxidase catalyzes the last step of biosynthesis of phytohormones indole acetic acid and abscisic acid. Xanthine dehydrogenase is involved in the catabolism of purines (Mendel and Haensch 2002; Kaiser *et al.* 2005). With the exception of bacterial nitrogenase, Mo-enzymes share a similar heterocyclic compound (pterin) composed of a pyrazine ring and a pyrimidine ring at their catalytic sites, the Mo cofactor. Mo is biologically inactive unless it is complexed with its cofactor. Among the micronutrients essential for the growth of plants and microsymbionts, Mo is required in minute amounts. In contrast to bacteria, no specific Mo uptake system is known in higher plants, but since molybdate (A molybdate is a compound containing an oxoanion with molybdenum in its highest oxidation state of 6) and sulfate behave similarly and have similar structure, uptake of molybdate could be mediated unspecifically by one of the sulfate transporters. Transport of Mo into different plant organs occurs via xylem and phloem. Pterin-bound Mo constitutes the cofactor of important plant enzymes involved in redox processes e.g. nitrate reductase, xanthine dehydrogenase, aldehyde oxidase and probably sulfite oxidase. Biosynthesis of the Mo cofactor (Moco) starts with a guanosine-X-phosphate. Subsequently, a S-free pterin is synthesized; then S is added and finally Mo is incorporated. In addition to the molybdopterin enzymes, small molybdopterin binding proteins without catalytic function are known to probably be involved in the storage of Moco (Williams and Frausto da Silva 2002; Sauer and Freibert 2003). In symbiotic systems N supply of the host plant is strongly influenced by the availability of Mo in the soil, since both bacterial nitrogenase and NADPH-dependent nitrate reductase of mycorrhizal fungi are Mo enzymes. In *Nicotiana tabacum* Mo had negative effect on nicotine content (Tso *et al.* 1973). Aziz *et al.* (2010) noticed significant increase in various plant growth parameters viz. and chemical composition by the use of varying concentrations of Zn (0,100 and 200 ppm) in *Cymbopogon citratus*. However, the greatest increase in plant height, fresh and dry weight yield was noticed by increasing Zn application and 200 ppm. However, Das *et al.* (2005) and Pandey *et al.* (2006) observed similar results in other crops of medicinal importance, which shows the regulation of metabolic activities governed by mineral elements.

B has been shown to be an essential element for higher plants in several metabolic processes in the plant (Blevins and Lukaszewski 1998; Reid *et al.* 2007). The requirement

of plants for B is as little as 2 ppm and it may be present in plant tissue up to 75 ppm. It is required for translocation of sugars and also regulates flowering and fruiting, cell division, salt absorption, hormone movement, pollen germination, carbohydrate metabolism, H₂O use and N assimilation in the plants (Brown *et al.* 2002; Camacho-Cristobal *et al.* 2008). Evidence in the literature supports the idea that the major functions of B in growth and development of plants are based on its ability to form complexes with the compounds having *cis*-diol configurations (Hu *et al.* 1997). The formation of B complexes with the constituents of cell walls and plasma membranes as well as with the phenolic compounds seems to be a decisive step affecting the physiological functions of B. B seems to be of crucial importance for the maintenance of structural integrity of plasma membranes (Hu *et al.* 1997). This function of B is mainly related to stabilization of cell membranes owing to its association with the membrane constituents. B also protects plasma membranes against the oxidative damage caused by the ROS. In B-deficient plants, plasma membranes become highly leaky and lose their functional integrity. Enhanced oxidation of phenols is responsible for generation of reactive quinones, which subsequently produce extremely toxic O₂ species, thus, increasing the risk of oxidative damage to vital cell components such as membrane lipids and proteins. In B-deficient tissues, enhancement in levels of toxic O₂ species may also occur as a result of impairments in photosynthesis and antioxidative defense systems. Recent evidence shows that the levels of ascorbic acid, non-protein SH-compounds (mainly glutathione) and glutathione reductase, the major defense systems of the cells against toxic O₂ species, are reduced in response to B deficiency. There is also increasing evidence that, in the heterocyst cells of cyanobacteria, B is involved in protection of nitrogenase activity against O₂ damage. Boric acid may bind to *cis*-diols, i.e. the compounds that contain pairs of *cis* hydroxyl groups such as sugar and sugar alcohols. The effects of B on plant growth and development depend on its specific complexing with borate and polyhydroxy substrates, enzyme protein and the co-enzymes with proper configuration of OH groups (Blevins and Lukaszewski 1998; Herrera-Rodríguez *et al.* 2010). However, meager studies on the role of B in metabolism of medicinal plants have been conducted in recent years which need special attention to have a better understanding.

4. Cadmium and cobalt

Cd and Zn are chemically very similar. Cd is able to mimic the behavior of essential element Zn in its uptake and metabolic function. Unlike Zn, Cd is toxic both for plants and animals (Sandalio *et al.* 2001; Akguc *et al.* 2008). The basic cause of toxicity probably lies in its higher affinity with thiol group (SH) of enzymes and other proteins. In many plants, excess Cd may also disturb Fe metabolism and can cause chlorosis of plant parts (Kosma *et al.* 2004). Edelbauer and Stangl (1992) found a direct effect of Cd in reducing element uptake; the toxicity of this heavy metal affected various enzymes and membrane systems, but indirect effects are also conceivable. The reduction of micronutrient levels in the case of Cd exposition appeared not only to depend upon the applied Cd concentration to the substrate but also on the micronutrient status of the plant (Lux *et al.* 2011).

Co, a transition element, is an essential component of several enzymes and co-enzymes (Aziz *et al.* 2007). It has been shown to affect growth and metabolism of plants to different degrees depending on the concentration and status of Co in the rhizosphere and soil. Co interacts with other elements to form complexes. The cytotoxic and phytotoxic activities of Co and its compounds depend on the physico-chemical properties of these complexes, including their electronic structure, ion parameters (charge-size relations) and coordination. Thus, the competitive absorption and mutual activation of associated metals influence the action

of Co regarding various phytochemical reactions. Co is not found at the active site of any respiratory chain enzymes (Atta-Aly *et al.* 1991). There are two sites of action of Co^{2+} in mitochondrial respiration, since it induces different responses towards different substrates such as alpha-keto glutarate and succinate. In lower organisms, Co^{2+} inhibits tetrapyrrole biosynthesis, but in higher plants it probably participates in chlorophyll *b* formation (Gad and Kandil 2009). The exogenously added metal causes morphological damage to plastids and brings about changes in the chlorophyll content (Gad and Kandil 2009; Tan *et al.* 2010). It also inhibits the differentiation of starch grain and alters the structure and number of chloroplasts per unit area in the leaf (Gad and Kandil 2009). The role of Co in photosynthesis is controversial. Its toxic effect results in the inhibition of PS-II. It inhibits either the function of reaction centre or that of component of PS-II acceptor by modifying the electron acceptor site of secondary quinone (QB). Co^{2+} reduces the export of photoassimilates and the extent of dark fixation of CO_2 (Kurosaki 1996). In C₄ plants, it hinders fixation of CO_2 by inhibiting the activity of the enzymes involved (Palit and Sharma 1994; Gad and Kandil 2009). Relatively higher concentrations of Co are toxic to the plants (Gal *et al.* 2008). Toxic effects of Co on plant morphology include leaf fall, inhibition of greening, discolored veins, premature leaf closure and reduction in shoot weight. Being component of vitamin B-12 and cobamide coenzyme, Co^{2+} helps in the fixation of molecular N in root nodules of leguminous plants (Yadav and Khanna 2002; Bolachander *et al.* 2003; El-Sheekh *et al.* 2003; Parmar and Chanda 2005; Gad 2006; Jayakumar *et al.* 2008). However, in cyanobacteria, CoCl_2 inhibits the formation of heterocyst, ammonia (NH_3) uptake and nitrate reductase activity. The interaction of Co with other metals mainly depends on the concentration of the metals used. For example, high level of Co^{2+} induces Fe deficiency in plants and suppresses uptake of Cd by roots (Jayakumar and Jaleel 2009). It also interacts synergistically with Zn, Cr and Sn (Vinay *et al.* 1996; Tan *et al.* 2010). The beneficial effects of Co include retardation of senescence of leaf, increase in drought resistance in seeds, regulation of alkaloid accumulation in medicinal plants and inhibition of ethylene biosynthesis (Gad and Kandil 2009). The plant mechanism of resistance to toxic concentration of Co may be associated with its intracellular detoxification rather than the defective transport. Among higher plants, only few advanced Cu-tolerant families showed cotolerance to Co^{2+} . Tolerance of plants against Co^{2+} may sometimes determine the taxonomic shifting of several members of the Nyssaceae. Due to the high Co content in serpentine soil, the uptake of essential elements by plants is reduced, a phenomenon known as "serpentine problem," for New Caledonian families like Flacourtiaceae (Yang *et al.* 1985). A high amount of soil Ca may neutralize the toxic effects of heavy metals in the adaptable genera grown in this type of soil. The biomagnification of potentially toxic elements, such as Co, coming from coal ash or H_2O into food webs, needs additional studies for effective biological filtering (Palit and Sharma 1994). However, Co behaves like other heavy metals such as Fe, Mn, Zn and Cu, which tend to form chelate compounds. Additionally, Co is essential for symbiotic N₂ fixation (Ahmed and Evans 1960). In *Nicotiana*, application of Co increased the concentration of N, P, K and Na, while the concentration of Mg decreased (López-Lefebre *et al.* 2002). Ca also showed a positive response to Co application; Fe, Mn, Cu and Zn also showed positive response to Co application, but at lower concentrations. Similarly, Co and Ca content in the soil were positively correlated with seed alkaloid content of *Colchicum autumnale*, which was probably due to enhanced uptake of various other minerals that significantly elevated metabolic activities in the plant (Poutaraud and Girardin 2005; Aziz *et al.* 2007).

5. Aluminum, nickel, chromium and selenium

More than 15% of the earth's crust is made up of Al_2O_3 .

The solubility of Al in soil is very low, due to which neutral and alkaline soils may not be toxic to plant growth. There are some evidences that Al in low concentration is beneficial for plant growth though the mechanism is not clear (Delhaize and Ryan 1995; Matsumoto 2000; Vardar *et al.* 2006). Higher plants usually contain Al up to 200 ppm in the dry matter. Generally, Al content in roots is much higher than in the upper plant parts. It is supposed that a substantial proportion of root Al is fixed in free space. The dissolution of Al hydroxy compounds in soil depends much on soil pH. Low soil pH values may result in high level of soluble Al, which is toxic to plants (Ozyigit and Akinci 2009). The first observable effect of Al on plants is limitation in root growth (Jones and Kochian 1995). Frequently, phosphate uptake and phosphate translocation to the upper plant parts are affected. In the plant cell, Al may interfere with the phosphate metabolism through the formation of stable Al-phosphate complexes. According to Siegel and Haug (1983), Al binds to calmodulins and may interfere with various enzymatic processes. The plasma membrane is also affected by high Al concentrations (Vardar and Ünal 2007). Al may have a detrimental effect on ion uptake. Grimme (1983) noticed that Al especially retards the uptake of Mg^+ (Jones and Kochian 1995).

Ni is closely related to Co both in its chemical and physiological properties. It readily forms chelate compounds and can replace other heavy metals from physiologically important sites. High Ni concentration has a toxic effect on plants. Its high concentration in nutrient medium reduces the uptake of most other nutrients (Cooke and Inkson 1955). Knight and Cooke (1956) noted a reduction in nutrient uptake caused by the damaging effects of high Ni concentration on the roots. Nonetheless, Ni has shown to be an integral part of the enzyme urease isolated from jack bean (*Canavalia ensiformis*) seeds (Brown *et al.* 1990). The biological significance of Ni as a possible micronutrient has earlier been reviewed by Aziz *et al.* (2007).

There is no evidence of any essential role of Cr in plant metabolism. Soil Cr is largely unavailable to plants because it occurs in relatively insoluble compound such as chromite $\text{Fe-Cr}_2\text{O}_4$ or mixed oxides of Cr, Al and Fe. Besides, it is found in silicate lattices. The rate of uptake and translocation of Cr by plants is low. It is supposed that Cr^{3+} and CrO_4^{2-} are taken up by two different mechanisms (Bolland 1983). Uptake of CrO_4^{2-} is depressed by SO_4^{2-} . Cr is found to inhibit the uptake of other minerals such Fe, Mn, Cu and Zn in *Phyllanthus amarus* and *Solanum nigrum* (Rai *et al.* 2007).

Selenium (Se) resembles with S in its chemical properties. In its uptake, there occurs a competitive effect between selenate and sulphate, indicating that both the ions have affinity for the same carrier sites (Antal *et al.* 2010). The incorporation of Se into amino acids analogous to those of S has also been observed in a number of plant species (Paterson and Butler 1962). It is believed that Se compounds interfere with S metabolism through replacement. However, Se is not able to replace S in all its metabolic functions. The differences between plants in their ability to accumulate and tolerate Se have not been fully explained. It is reported that in non-accumulator plants, Se remains more in proteins (Vogrinic *et al.* 2009; Antal *et al.* 2010).

REGULATORY ROLE OF MINERAL ELEMENTS IN PROTEIN SYNTHESIS

Major steps in protein synthesis

All the metabolic activities in living organisms are catalyzed by enzymes, which are proteins. Cells are capable of synthesizing the proteins, essential for the modulation and maintenance of cellular activities. Formation of proteins, using amino acids (building blocks), is based on the information encoded in the nucleic acids (Deoxyribose nucleic acid (DNA) and Ribose nucleic acid (RNA)). Protein synthesis generally consists of two major steps: transcription

and translation (Berg *et al.* 2002; Meiger and Thomas 2002). Proteins are synthesized in the cytoplasm according to the information present in the messenger RNA (Wickens *et al.* 2000; Mendez and Richter 2001). Transcription is the process, in which the genetic information present in the DNA is used to produce a complementary RNA strand. One of the strands of DNA double helix is used as a template by RNA polymerase to synthesize a messenger RNA (mRNA). The mRNA migrates from the nucleus to cytoplasm, where it is free to move. The mRNA goes through different types of maturation steps including the one called mRNA splicing, through which the non-coding nucleotide sequences are eliminated. The coding mRNA sequence is a unit of three nucleotides called a codon (Gnatt *et al.* 2001). During translation, the mRNA molecules bind to protein-RNA complexes called ribosomes located in the cytosol (Lodish *et al.* 2000; Kapp and Lorsch 2004). The ribosome binds to mRNA at the start codon (AUG) that is recognized only by the initiator tRNA where they are translated into polypeptide sequences (Nika *et al.* 2000; Pestova *et al.* 2001; Schmitt *et al.* 2002; Sonenberg and Hinnebusch 2009). The ribosome mediates the formation of a polypeptide sequence based on the mRNA sequence. During this stage, complexes, composed of an amino acid linked to tRNA, sequentially bind to the appropriate codon in mRNA by forming complementary base pairs with the tRNA anticodon. The ribosome moves from codon to codon along the mRNA. Amino acids are added one by one and are thus translated into polypeptidic sequences as directed by DNA and represented by mRNA. At the end, a release factor binds to the stop codon, terminating the translation step and releasing the complete polypeptide (protein) from the ribosome (Nissen *et al.* 2000; Valadkhan and Manley 2001; Schmitt *et al.* 2002; Mandal *et al.* 2003; Bruce 2008).

N is by far the largest N fraction and amounts to about 80-85% of the total N present in proteins present in green plants (Kusano *et al.* 2011). The N of nucleic acids makes about 10% and soluble amino N about 5% of total N present in plant material. In vegetative plant parts, the enzymes constitute the major portion of proteins, whereas in seeds and grains the major protein fraction is made up of storage protein. Besides, N is also an essential constituent of various coenzymes. Glutamate and glutamine are the first two amino acids synthesized during the process of NH₃ assimilation. Glutamate and aspartate as well as their amides accumulate when high levels of inorganic N are applied to plants. Under such conditions the excess inorganic N is obviously used in protein synthesis (Coruzzi and Bush 2001; Coruzzi 2003; Schachtman and Shin 2007; Krouk *et al.* 2010).

S is essential for the conversion of N into amino acids and the linkage of these amino acids into complete proteins. S accounts for about 1% of the plant tissue. S deficiency causes accumulation of nitrates in plant tissue instead of forming amino acids and protein (Schnug and Haneklaus 1998; Kertesz and Mirleau 2004). Elemental S is oxidized into a plant-useable form of sulphate by an S-oxidizing microorganism known as *Thiobacillus*. This important soil organism functions actively when the soil is well aerated. Compact soils create anaerobic conditions that can reduce sulphates to toxic acids of sulfur and gases. Amino acids are the basic structural units of all proteins and they play a very important role in the manufacturing of proteins. There are two amino acids which contain an S atom and are called S-containing amino acids; viz. cysteine and methionine (Scherer 2001; Ostrowska *et al.* 2008). Cysteine is an S-containing amino acid and is closely related to cystine. Cystine consists of two cysteine molecules joined together. Cysteine contains a sulphhydryl (-SH) group; this is tremendously reactive and can form H bonds. This S-containing amino acid is very important because it can also form disulphide bridges. S-containing amino acids like cysteine are required for the production of taurine and are also a component of the antioxidant glutathione. Methionine is another S-containing amino acid; it is the "start" amino acid in the process of

protein synthesis, and is therefore used as beginning step of every single protein formed. Its S atom carries thioether linkage and is relatively unreactive (Scherer 2001).

The unique function of phosphate in metabolism is its formation of pyrophosphate bond, which allows energy transfer. Uridine triphosphate (UTP), cytidine triphosphate (CTP) and guanosine triphosphate (GTP) are analogous compounds to ATP. Uridine triphosphate is required for the synthesis of sucrose and callose, CTP is required for the synthesis of phospholipids, and GTP is required for the formation of cellulose. All of these nucleotide triphosphates (ATP, UTP, GTP and CTP) are also involved in the synthesis of ribonucleic acid (RNA). For the synthesis of deoxyribonucleic acid (DNA), deoxy form of the nucleotide triphosphates is required. In the deoxy form, ribose sugar of the nucleotide is substituted by deoxyribose sugar. The phosphate group in nucleic acid bridges the ribose (RNA) or deoxyribose (DNA) sugars with another ribose or deoxyribose sugar by two ester bonds. DNA is the carrier of genetic information while various forms of RNA function in protein synthesis (Berg *et al.* 2002; Meiger and Thomas 2002; Noller *et al.* 2002).

Generally, when plants are Mg-deficient, a proportion of non-protein N increases, indicating that Mg deficiency inhibit protein synthesis. This does not result from the lack of synthesis of particular amino acids, as it is in the case of S deficiency. The effect is probably caused by dissociation of the ribosomes into their sub-units in the absence of Mg²⁺ (Watson 1965). Thus, it appears that Mg²⁺ is used to stabilize the ribosomal particles. This ribosomal configuration is necessary for protein synthesis. Mg is also believed to have a similar stabilizing effect on the matrix of the nucleus. According to Wunderlich (1978) binding of ribosomal sub-units is achieved by a bridging effect of Mg²⁺ on neighbouring indiffusible anion. The transfer of amino acyl from amino acyl tRNA to the polypeptide chain is also probably facilitated by Mg²⁺. High concentrations of MgSO₄ are most favorable for both root growth and ginsenoside accumulation (up to 8.89 mg/g dry weight (DW)) in *Panax ginseng* (Yu *et al.* 2001).

B is essential in the synthesis of the nitrogenous base uracil, which is needed for RNA metabolism (Albert 1968). The distribution of Co in plants is entirely species-dependent and its uptake is controlled by different mechanisms in different species (Moreno-Caselles *et al.* 1997a, 1997b). Toxic concentrations of Co inhibit active ion transport. In higher plants, absorption of Co²⁺ by roots involves active transport. Its transport through cortical cells is operated by both passive diffusion and active process. In the xylem, the metal is mainly transported by the transpirational flow. Its distribution through the sieve tubes is acropetal; it takes place by complexing with organic compounds. The lower mobility of Co²⁺ in plants restricts its transport to leaves from stems (Page and Feller 2005). It also plays a vital role in DNA synthesis by reducing ribonucleotide to deoxyribonucleotide. It is essential for production of methionine synthase, which is involved in protein synthesis. High concentrations of Co hamper RNA synthesis, and decrease the amounts of DNA and RNA in the cell probably by modifying the activity of a large number of endo- and exo-nucleases (Bakkaus *et al.* 2005; Khan and Khan 2010). Investigations indicated that Al is adsorbed on DNA double helix and thus inhibits the separation of its two strands. The effect of NaCl was elucidated by Niknam *et al.* (2006) on a medicinally important plant *Trigonella foenum-graecum* L. and *Trigonella aphanoneura* Rech. The protein contents in both of these species were significantly higher in NaCl treated plants compared to the untreated plants. Regarding Mo, there is an important finding reported by López *et al.* (2007), which depicted that the dry matter production and protein content in red clover increased significantly as Mo concentration increased in the shoots from 0.5 to 1.0 mg Mo kg⁻¹ dry matter. Ma *et al.* (2010) studied the effect of lanthanum nitrate (La³⁺) and cerium nitrate (Ce⁴⁺) on *Echinacea angustifolia*, an important medicinal plant. They

noticed the dose-dependent effects of La^{3+} on soluble protein content, peroxidase activity and shoot differentiation. Yin *et al.* (2006) noticed that growth of *Catharanthus roseus* cells in suspension-culture ceased during phosphate starvation, but these cells grew up again upon addition of P_i even after long-term starvation. They studied the metabolic fate of [^{32}P] P_i in 1-week-old stationary phase cells in ordinary culture and in 1- or 2-week-old P_i -starved cells. Immediately after P_i administration, the most heavily labeled organic compounds were nucleotides, followed by sugar phosphates. P_i starvation for 2 weeks slowed the speed of incorporation of ^{32}P into nucleotides. The RNA, protein, and free nucleotide contents decreased during P_i starvation; however, these compounds, especially the nucleotides, increased markedly in 24 h after the addition of P_i . These responses were found in all the cells, although total amounts of these compounds were lower in long-term P_i -deficient cells. Of the nucleotides, a marked increase was observed in nucleoside triphosphates and UDP-glucose. The transcript level of phosphate transporter and the activities of acid phosphatase, 5'- and 3'-nucleotidase, and adenosine nucleosidase were reduced by the addition of P_i . In contrast, the activities of adenine phosphoribosyltransferase, nicotinate phosphoribosyltransferase and nicotinamidase, which are salvage enzymes of purine and pyridine nucleotides, were markedly increased in P_i -fed cells. Little or no increase was observed in adenosine kinase.

REGULATORY ROLE OF MINERAL ELEMENTS IN ION EXCHANGE

Every living cell acquires raw materials (especially minerals nutrients) from its surroundings for the biosynthesis of energy and release of the byproducts of metabolism in the environment. Some non polar compounds might be dissolved in lipid bilayer and cross the membrane unassisted by any carrier or transporter, while charged or polar compounds or ions require a membrane protein (carrier or transporter) for their transportation across the membrane. However, in some cases a membrane protein simply facilitates the diffusion of a solute down its concentration gradient, but such a transport often occurs against a gradient of concentration, electrical charge, or both; in this case, the solute is pumped in by a process that requires energy (Chrispeels *et al.* 1999). The energy is provided by the hydrolysis of ATP or may it be supplied during the movement of another solute down its electro chemical gradient with enough energy to carry yet another solute up its gradient; the ions may also move across the membrane through ion channels formed by proteins, or they may be carried across by ionophores, small molecules that mask the charge of ions and allow them to diffuse through the lipid bilayer (Rubio *et al.* 1995; Gassmann *et al.* 1996; Sze *et al.* 1999). Except the few exceptions, the translocation of small molecules across the plasmas membrane is mediated by proteins such as transmembrane channels, carriers or ion pumps (Chrispeels *et al.* 1999; Lalonde *et al.* 1999; Sze *et al.* 1999). Within the eukaryotic cell, different compartments have different concentrations of metabolic intermediates, products and ions, and these substances move across the intracellular membranes through tightly regulated, protein-mediated processes.

Nitrate is an important macronutrient. It is also acts as a signal for plant growth. Its level in the soil may vary by three to four times of its magnitude. Consequently, plants have evolved regulated energy dependent systems for the uptake of nitrate, using both high and low affinity transporters (Chrispeels *et al.* 1999). Most plant species are able to absorb and assimilate nitrate (NO_3^-), ammonium (NH_4^+), urea and amino acids as N sources, but the response to a particular form of N varies from species to species. In general, most crop plants prefer a mixture of NH_4^+ and NO_3^- and take up a higher proportion of NH_4^+ present in the soil solution only in specific cases, as in the case of *Papaver somniferum*. Variation in soil nitrate level affects the rate of plant

growth and concentration of NO_3^- stored in plants. In unlimited supplies of NO_3^- , root and shoot concentrations of NO_3^- can reach up to 100 mM, most of which is stored within the vacuoles. However, the cytoplasm is a more important compartment for NO_3^- metabolism (Chrispeels *et al.* 1999). In soil, NO_3^- is carried towards the root by bulk flow and is absorbed into the epidermal and cortical symplasm. Within the root symplasm, NO_3^- has the four following fates: (1) reduction to NO_2^- by the cytoplasmic enzyme nitrate reductase; (2) efflux back across the plasma membrane to apoplasm; (3) influx and storage in vacuole; or (4) transport to xylem for long-distance translocation to the leaves. Following long-distance translocation, NO_3^- must leave the xylem and enter the leaf apoplasm to reach the leaf mesophyll cells, where NO_3^- is again absorbed and either it is reduced to NO_2^- or stored in the vacuole (Glass and Siddiqi 1995; Crawford and Glass 1998).

On reaching the root surface, uptake of NO_3^- into the cell requires energy, even though external NO_3^- concentration remains in low range. However, in spite of expenditure of energy on NO_3^- uptake, there appears to be a significant efflux of NO_3^- across the plasma membrane. The rate of efflux increases with increasing external NO_3^- concentration. The NO_3^- efflux is passive, and there is evidence that it is saturable and NO_3^- -selective. Employing the use of inhibitors of RNA and protein synthesis has indicated that it is NO_3^- -inducible. However, nothing is known about the proteins involved in the efflux pathway (Chrispeels *et al.* 1999). The energy for ion uptake is provided by the proton gradient or pmf, which, based on typical cell parameters, is adequate to drive active NO_3^- uptake over a wide range of external NO_3^- concentrations. In addition, NO_3^- uptake is associated with depolarization (an increase in the positive charge inside the cell) of the plasma membrane. Physiological studies and molecular investigations in *Nicotiana* species and *Arabidopsis* indicated that several N forms, including NO_3^- , NH_4^+ and amino acids, may participate in this down regulation, which can occur at the mRNA level. Such a regulation is important for coordinating the root uptake as per shoot demand for N during the growth cycle of the plant. A common explanation, given for the mechanism of rapid uptake of NH_4^+ and inhibition of that of NO_3^- , is that NH_4^+ depolarizes the plasma membrane and, thus, reduces pmf for active NO_3^- uptake by $2\text{H}^+/\text{NO}_3^-$ symport mechanism (Taylor and Bloom 1998). This explanation is supported by the observations that indicate that NH_4^+ induces a sustained membrane depolarization and that the more depolarized the membrane potential is, the lower is the NO_3^- -induced current (Day *et al.* 2001; Roberts and Tyerman 2002; Kabała *et al.* 2003). However, since K^+ is also known to depolarize the membrane electrical potential without an equivalent reduction in NO_3^- uptake, the reduction in electrical potential alone seems insufficient to explain the short term inhibition by ammonium (García-Sánchez *et al.* 2000). An increase of N supply in *Catharanthus roseus* stimulated K and P uptake and their translocation to the leaves has been proven by the results of the chemical analysis (Lata and Sadowska 1996). El-Sherbeny *et al.* (2007) demonstrated the effect of different doses of N in the form of compost on an important medicinal plant *Ruta graveolens* L. They found positive effect of the compost doses on the amount of essential oil, rutin, coumarin and other secondary metabolites. Pavlíková and Thlustoš (1994) concluded that it is also important to choose a suitable form of N fertilizer, especially when the uptake of N from soil and the seed yields are concerned. The need for an optimal supply of N for poppy plants begins shortly after germination and lasts till the stage of formation of generative organs.

Potassium channels

The translocation of cations across biological membranes is an inherent feature associated with numerous physiological processes, including growth and development, signal transduction and cell homeostasis. It is well known that adeno-

sine and/or guanosine 3', 5'-cyclic monophosphate (cNMP; cAMP or cGMP) are important secondary messenger signaling molecules in eukaryotic and prokaryotic cells. They are typically involved in transduction of a signal into a specific cellular response. Cyclic nucleotide gated cation channels (CNGCs) were first identified in animals, where they are involved in visual, gustatory and olfactory signal transduction as well as in other physiological processes. Although plant CNGCs showed relatively overall low sequence identity with animal CNGCs, they do share significant homology at their pore regions and cyclic nucleotide binding sites (CNBSs). The functional channel is most likely represented as a homo-tetramer, although heterotetrameric channels, organized in situ, should not be ruled out. Each subunit consists of six transmembrane domains (S1–6) containing a pore-forming region, with lower hydrophobicity, between S5 and S6 region. Moreover, CNGCs includes a CNBS intracellularly located downstream from the pore. These channels are directly gated upon binding with cAMP or cGMP and are permeable to a variety of monovalent cations as well as to Ca^{2+} . In addition to these characteristics, plant CNGCs include a putative Ca^{2+} /calmodulin binding site (CaMBS) embedded within the CNBS (Leng *et al.* 1999). Ca^{2+} /calmodulin ($\text{Ca}^{2+}/\text{CaM}$) and cNMPs are essential components in a number of well characterized signaling pathways, strengthening the argument that plant CNGCs play critical roles in Ca^{2+} signal transduction (Merclier and Berkowitz 2002).

K^+ (K^+) is required for every major step of protein synthesis. The "reading" of the genetic code in plant cells to produce proteins and enzymes that regulate all growth processes, would be impossible without adequate K^+ . When plants are deficient in K^+ , proteins are not synthesized despite an abundance of available N. Instead, the protein "raw materials" (precursors) such as amino acids, amides and nitrate are accumulated. The enzyme nitrate reductase (NR) catalyzes the formation of nitrite from the nitrate that, in turn, gets converted to ammonium ions after being acted upon by nitrite reductase, leading to the formation of proteins (ammonia assimilation). Since K^+ is likely responsible for the activation and synthesis of the first enzyme (NR) in the series, NR is also thought to be responsible for protein synthesis apart from other important enzymes. K^+ is a major nutrient in higher plants, where it plays a role in turgor regulation, charge balance, leaf movements and protein synthesis. Terrestrial plants are able to sustain growth at micromolar external concentrations of K^+ , at which K^+ uptake across the plasma membrane of root cells could be energized despite the presence of a highly negative membrane potential. However, the mechanism of such an energized K^+ uptake has long been remained obscure. Therefore, whole-cell mode patch, clamping [a technique for the study of measurement of membrane potentials and electrical properties of plant cell (Miedema and Assmann 1998)] has been applied to root protoplasts by *Arabidopsis thaliana* to characterize the membrane currents resulting from the application of micromolar K^+ concentrations. Analysis of whole cell current/voltage relationships in the presence and absence of micromolar K^+ concentrations enabled direct testing of K^+ transport for possible energization by cytoplasmic ATP and the respective trans-membrane gradients of Na^+ , Ca^{2+} , and H^+ . Subtracted current/voltage relations for K^+ -dependent membrane currents are independent of ATP and could be reverse at potentials that imply H^+ -coupled K^+ transport with a ratio of 1 $\text{H}^+:\text{K}^+$. Furthermore, the reversal of the potential of K^+ current shifts to negative mode as the external H^+ activity is decreased. The K^+ -dependent currents saturate in micromolar concentration range with an apparent K_m of 30 mM, a value that is in close agreement with previously reported K_m values for high-affinity K^+ uptake. The high-affinity K^+ uptake in higher plants is mediated by a $\text{H}^+:\text{K}^+$ symport mechanism, which is competent in driving K^+ accumulation to equilibrium ratios in excess of 10⁶-fold (Maathuis and Sanders 1995).

K^+ (K^+) is the most abundant cation in plants, whereas closely related Na^+ (Na^+) ion is toxic to most plants at high millimolar concentrations. Both K^+ deficiency and Na^+ toxicity are major constraints to crop production worldwide. In fact, K^+ counteracts Na^+ stress, while Na^+ , in turn, can alleviate K^+ deficiency up to a certain degree (Pascal *et al.* 2002). The beneficial effect of K^+ on photophosphorylation has been observed by Watanabe and Yoshida (1970), Hartt (1972) and Pfluger and Mengel (1972). Overnell (1975) and Weller and Hofner (1974) have reported that K^+ stimulates photosynthetic O_2 production. These observations supported the view that K^+ has direct influence on electron transport in the photosynthetic ETC. It might be supposed that the movement of K^+ from the thylakoid spaces into the stroma of chloroplasts, which occurs in light, depolarizes the thylakoid membrane that, in turn, favours the electron flow in transport chain (Mitchell 1966). Depolarization of plasma membrane by K^+ uptake has a direct influence on phloem loading. Numerous researchers have shown that K^+ enhances the translocation of photosynthates. K^+ not only promotes the translocation of newly synthesized photosynthates, but also has a beneficial effect on the mobilization of stored material. Liu and Zhong (1996) noticed the effects of initial K^+ concentrations (0–60 mM) on the kinetics of cell growth, nutrients metabolism and production of ginseng saponin and polysaccharide in suspension cultures of *Panax ginseng* by altering KNO_3 and NaNO_3 concentrations at 60 mM of total NO_3^- . They showed that comparatively a higher concentration of soluble sugars was stored within the cells under K^+ -deficiency, and that there was a curvilinear relationship between initial K^+ concentration and active biomass accumulation. The K^+ consumption by the cells was dependent on the initial medium K^+ concentration and there was a linear correlation between K^+ consumption and nitrate consumption. Although K^+ had little effect on the content of ginseng polysaccharide, the saponin content was remarkably enhanced with the increase in initial K^+ concentration within a range of 20–60 mM.

Calcium gradient and oscillations in growing pollen tubes

Pollen tube growth delivers two sperm cells to embryo sac; it is essential for sexual reproduction in higher plants (Feijó *et al.* 2001; Hepler *et al.* 2001; Cárdenas *et al.* 2008). The process has important and unique features and is extremely fast. For example, the rate of movement of sperm cells is 1 cm per hour in corn. The process is highly polarized and pollen tube growth is mainly confined to the tip ("tip-growth"). It possesses a guidance mechanism that determines the direction of growth of the tube (Parton *et al.* 2003; Wilken and Hepler 2007). Several aspects of tip-growth of the pollen tube have been deciphered; for example, the Golgi apparatus produces vesicles containing cell wall precursor, and through cytoplasmic streaming the vesicles flow to the apex of the pollen tube, where they fuse with the plasma membrane and secrete their contents into the cell wall. It is unclear how these events are orchestrated to achieve the high degree of polarity during pollen tube growth, although recent work indicated that different ions, especially Ca ions and protons play an important role in this regard (Feijó *et al.* 2001; Hepler *et al.* 2001; Song *et al.* 2009).

It is well known that both Ca ions and protons are essential for growth (Hepler *et al.* 2001; Iwano *et al.* 2009; Song *et al.* 2009). Ca ions must be present in the growth medium at a concentration above 10 μM , but these ions have detrimental effect above 10 mM. A high proton concentration or low pH facilitates both pollen germination and pollen tube growth. Recent work, focused on the intracellular and extracellular status of Ca ions and protons has been conducted, using the technology that is available since the last decade. Studies on intracellular Ca, conducted by using fluorescent indicator dyes (Holdaway-Clarke and Hepler 2003), or the photoprotein and aequorin (Messerli *et al.*

al. 2000), revealed that the growing pollen tube possessed a highly focused gradient of free Ca at its extreme apex (Song *et al.* 2009). The intracellular Ca concentration, immediately adjacent to the plasma membrane, remains 2,000 to 10,000 nM (Messerli *et al.* 2000), but it declines abruptly to a basal level i.e. 200 nM or less at 20 μ m distance from the pollen tube tip. Recent studies showed that the apex of pollen tube contains an acidic region with a large alkaline domain towards the base of the clear zone (Feijó *et al.* 1999). The pH gradient is large and extends from 6.8 at the tube apex to 7.8 at 20–30 μ m distances from the tip. The extracellular influx of Ca^{2+} and the intracellular Ca gradient, thus, mark the direction of pollen tube growth. Extracellular protons, like Ca ions, also show an influx at the apex of the growing pollen tube. However, in contrast to Ca ions, protons display a marked efflux along the side of the tube, close to the base of clear zone, in a position that corresponds to the location of the intracellular alkaline band (Feijó *et al.* 1999; Ikeda *et al.* 2003; Watahiki *et al.* 2004). Vernoica *et al.* (2002) have previously noticed that the increases in cytosolic free Ca^{2+} triggered by the self-incompatibility (SI) response in incompatible *Papaver rhoesas* pollens. In a recent study, they used an ion-selective vibrating probe to measure changes in the extracellular Ca^{2+} fluxes around poppy pollen tubes. They confirmed that there was an oscillating Ca^{2+} influx directed at the apex of the pollen tube and that Ca^{2+} influx also occurred at the shanks of pollen tubes. Additionally, as per the challenge with self-incompatibility (SI) proteins, there is a stimulation of Ca^{2+} influx along the shank of incompatible pollen tubes, approximately 50 mm behind the pollen tube tip. Thus they demonstrated that SI-induced Ca^{2+} influx played a role for influx of extracellular Ca^{2+} in the SI response. In a study on medicinally important plant *Nicotiana tabacum*, Tuna and Burun (2002) concluded that all heavy metals have negative effects on pollen characteristics; however, the damage to pollen varies with the dose. All plants showed some tolerance against different pollutants.

The genus *Crataegus* (hawthorn) includes some important species having medicinal, ornamental and sanitary properties. Among them, *Crataegus douglasii* (black hawthorn) and *Crataegus oxyacantha* (red hawthorn) are mostly used, having more nutrients, medical, ornamental and sanitary properties and used as a main dry tolerant rootstock recently. The fruits are the part having commercial value and the most important factors involved in fruit set are pollination, pollen tube growth and fertilization. Sharafi (2010a) studied pollen germination and tube growth of some East-Azerbaijan, Iran indigenous hawthorn genotypes in different *in-vitro* media for receiving the suitable medium for pollen germination and identifying favorable genotypes for using in the future breeding and orchard establishment programs. The *in-vitro* medium contained different concentrations of sucrose (5, 10, 15 and 20%), boric acid (0.005, 0.01 and 0.02%) with (1.2%) agar and pollens were cultured in different media and maintained about 24 h at 22°C but growth was stopped with raising chlorophorm. The study showed that the best *in-vitro* medium for hawthorn pollen germination and tube growth were composed of 15% sucrose, 0.005–0.01% boric acid and 1.2% agar and significant differences observed between various media in the studied pollen traits. Sharafi (2010b) also studied pollen germination and tube growth in some *Hashtroud* and *Maragheh* indigenous genotypes of *Rosa canina* as these were the main factors affecting fruit set. The pollens were cultured in an *in-vitro* medium containing 15% sucrose, 0.01% boric acid and 1.2% agar and maintained in 24°C in controlled condition. After 24 h pollens germination and growth were stopped with chlorophorm. The study concluded significant differences between germination percentage (PGP) and pollen tube length (PTL) of genotypes.

REGULATORY ROLE OF MINERALS IN ALKALOID BIOSYNTHESIS

Before discussing the regulatory role of minerals on alkaloid biosynthesis, it is necessary to describe the general information about alkaloids. Alkaloids are compounds of non-peptidic origin that typically contain N and have complex ring structures. They are produced primarily in higher plants but also in lower organism and in some animals having pharmacological importance. The name was derived from the word 'alkaline' in 1819 by Carle Meissner, while originally the term was used to describe any N-containing base (an amine in modern terms) having similar properties as basic salts derived from the alkali ashes of plants. The first identified alkaloid was morphine, from opium poppy (Luch 2009). Evidence suggests that alkaloids have been used by humanity as medicines and poisons for thousands of years. The first civilization that used them was probably the ancient Sumarians and Egyptians (Kutchan and Dittrich 1995). However, it was not until early nineteenth century that these compounds were reproducibly isolated and analyzed. Advances in analytical separation techniques, such as chromatography and mass spectroscopy, led to elucidation of the chemical structure of alkaloids (Southon and Buckingham 1989). When separated, in general they are colorless, crystalline solids, basic in nature and have definite melting points. Some exceptions are, however, known. For instance, some alkaloids are not basic and others are brightly colored or liquid. Still it is unclear as to why the plants produce alkaloids. Several theories have been proposed explaining the reason; of them some suggest that alkaloids are byproducts of normal plant metabolism or are produced as a means of defense against insects and animals or may be the reservoirs for molecules that plants often use. It is likely that all of these theories are correct up to some extent.

Classification of alkaloids

Alkaloids can be classified in three ways based on their chemical structure (nucleus containing N) or based on their biosynthetic pathway (the way they are produced in the plant) (Trease and Evans 1959). According to first type of classification as proposed by Hegnauer (1986), alkaloids are divided into three groups:

a. True alkaloids: The basic unit in biogenesis of true alkaloid is amino acid. They are poisonous and found in plants as salts of organic acids having heterocyclic ring with N. The non-N containing rings or side chains are derived from terpene units or acetate, while methionine is responsible for the addition of methyl groups to N atoms, e.g. morphine, codeine, etc. (Trease and Evans 1959; Aniszewski 2007).

b. Proto alkaloids: The basic unit in biogenesis of a proto alkaloid is also amino acid but they do not have heterocyclic ring with N. Instead, they may have N atoms outside the ring systems e.g., colchicine and ephedrine (Aniszewski 2007).

c. Pseudo alkaloids: The basic unit in biogenesis of proto alkaloid is terpenoids or purines instead of amino acids. They also have heterocyclic ring with N, e.g. caffeine, theobromine, and theophylline (Aniszewski 2007).

On the basis of biosynthetic pathways, alkaloids can also be divided into three groups (Trease and Evans 1959; Aniszewski 2007). Group I liberates amino N from the precursor during the biosynthesis of its alkaloids. Alkaloids belonging to this group are synthesized from diamino acid family, phenylalanine-tyrosine family and tryptophan family. They have a higher C:N ratio than their immediate precursors. In this process, amino acid is converted into alkaloids, liberating one molecule of ammonia. Group II includes those alkaloids, which have the same number of N atoms as

their precursor do. Group III includes those alkaloids, which have a lower number of C:N ratio compared to their precursor. This group is sometime called as the group of pseudo alkaloids e.g., steroid alkaloids, isoprene derivatives, etc. (Waller and Nowacki 1978).

The third type of classification system to be considered was offered by Manfred (2002). This classification concentrates on the N atom present within the chemical structure of the alkaloid, and classifies the alkaloids based on the environment of that N atom. If there are present multiple N atoms in the chemical structure of the alkaloid, the priority is given to the N atom present in the predominant ring. The following five groups of alkaloids may be framed based on this classification of alkaloids:

Heterocyclic alkaloids

The majority of the alkaloids occur in this group. These alkaloids have an N atom in a heterocyclic ring, or in some cases part of two heterocyclic rings (Trease and Evans 1959; Joule and Mills 2000).

i) Pyrrolidine alkaloids

Alkaloids of this group contain five N rings and are predominately synthesized from ornithine and lysine, e.g. hygrine, isolated from the leaves of *Erythroxylum coca* (coca plant); it is an important precursor to hyoscyamine and scopolamine (Trease and Evans 1959; Joule and Mills 2000).

ii) Terpenoid indole alkaloids

The indole alkaloids comprise a large family of alkaloids. Many of these natural products are physiologically active in mammals. For instance, the antimalarial drug quinine extracted from *Cinchona officinalis*, the antineoplastic drug camptothecin extracted from *Camptotheca acuminata*, the rat poison and homeopathic drug strychnine extracted from *Strychnos nux-vomica* and the antineoplastic chemotherapeutic agents vincristine and vinblastine extracted from *Catharanthus* (periwinkle) belong to this group of alkaloids. In fact, the chemical synthesis of these complex alkaloids would be of academic interest, but due to their low yields they are not likely to be applied commercially. The indole alkaloids have indole as their structural backbone and are derived from tryptophan. The indole system can be considered to consist of a pyrrole ring and a benzene ring fused together to form the two isomeric benzopyrroles: indole and isoindole (Trease and Evans 1959; Joule and Mills 2000).

The indole appears to be an essential constituent of the perfumes extracted from the flowers of some plants such as jasminum and citrus species. With respect to their structural features, indole alkaloids can be divided into two main classes: the first class of simple indole alkaloids and the second class of two structure-elements. That is, (1) tryptamine with indole nucleus and (2) C₉ or C₁₀ - monoterpane moiety, derived from secologanin.

A common example is ergine, one of the ergot alkaloids, isolated from many plants and fungi. Ergine is a hallucinogenic alkaloid having similar effects like indole alkaloids but has an overall lower level of visual stimuli (Laing 2003). Strychnine is another indole alkaloid, isolated from the *Strychnos nux-vomica* (strychnine tree) in Southeast Asia. It is used as a pesticide for killing small birds and rodents, as it is a very potent neurotoxin (Brams *et al.* 2011). The alkaloids of *Rauwolfia* and *Vinca* also belong to this group. For centuries, the natives of the Himalayan foothills are using the roots of *Rauwolfia* for healing afflictions ranging from snake bites to insanity. Reserpine is the most important constituent of *Rauwolfia*. Reserpine depletes adrenergic vesicles of their transmitters, thereby, reducing the amount of neurotransmitter released. This causes hypotension, bradycardia and increased gastrointestinal motility and increased the rate of defecation (Liles *et al.* 2006). The widespread reference in folklore of the oral hypoglycemic

properties of leaf extracts from the *Catharanthus roseus* prompted investigations which led to the discovery of dimeric indole-dihydroindole alkaloids i.e. vinblastine and vincristine. These alkaloids produced severe leukopenia in rats suggesting an anti-cancer potential especially for the treatment of leukemia (Zhigaltsev *et al.* 2005).

The indole alkaloids derived from tryptamine and secologanin can be classified according to the structural characteristics of their skeletons into the following eight types (Trease and Evans 1959; Joule and Mills 2000).

1. Corynanthean or C-type (sarpagine, yohimbine, ajmalicine)
2. Vincosan or D-type (vincoside)
3. Vallesiachotamon or V-type (vallesiachotamine)
4. Strychnan or S-type (vomicine)
5. Aspidospermatan or A-type (condylocarpine)
6. Eburnan or E-type (vincamine)
7. Plumeran or P-type (kopsine)
8. Ibogan or J-type (voaluteine)

iii) Pyrrolizidine alkaloids

The pyrrolizidine alkaloids have pyrrolizidine rings as their back bone, e.g. retronecine. Its ester derivatives are used as sex pheromones in lower animal species (Xiong *et al.* 2009).

iv) Pyridine alkaloids

Pyridine alkaloids have pyridine in their basic structure, e.g. nicotine. Nicotine, the addictive compound found in tobacco, can also be isolated from several other plants including *Solanum seethianum* (nightshade) (Lindvai and Vizi 2008). Nicotine interacts with nicotine acetylcholine receptors throughout the nervous system. In brain, by interfering with these receptors, nicotine increases the concentrations of several neurotransmitters including dopamine, which induces the feeling of relaxation (Dani and Bertrand 2007). Anabasine, an alkaloid found in *Nicotiana glauca* (tobacco tree), produces the same effects as nicotine. It also interferes with the nicotine acetylcholine receptors throughout the body (Lindvai and Vizi 2008).

v) Piperidine alkaloids

The piperidine alkaloids have a piperidine ring in their chemical structure, e.g. Coniine. In the history, the alkaloid coniine was used to kill Socrates. This alkaloid is found in *Conium maculatum* and *Sarracenia flava* (Graham 1997; Schep *et al.* 2009). Coniine is a potent neurotoxin and less than 0.2 g of its quantity can be fatal (Erickson 2005). It paralyzes the muscles by blocking nicotinic acetylcholine receptors on the post synaptic membrane of the neuromuscular junction. Eventually, paralysis moves into the respiratory system and subsequently the death occurs by asphyxiation (Schep *et al.* 2009). Lobeline, another piperidine alkaloid, is found mainly in members of the genus *Lobelia*. The alkaloid is used as a drug assistant to stop smoking and works through the stimulation of the release of dopamine and by blocking the reuptake of dopamine and serotonin (Dwoskin and Crooks 2002).

vi) Quinoline alkaloids

The quinoline alkaloids have quinoline rings in their chemical structure, e.g. quinine. It isolated from cinchona bark and has been used in treatment of malaria for centuries (PrayGod *et al.* 2008). It also has antipyretic properties. Its mechanism of action is by prevention of hemozoin biocrystallization inside the plasmodium, thus, causing a buildup of cytotoxic heme groups (Jones *et al.* 2007).

vii) Iso-quinoline alkaloids

The iso-quinoline alkaloids contain an iso-quinoline ring in their structure, e.g. papaverine found in *Papaver somniferum*.

rum (Shukla *et al.* 2010). Papaverine is a compound used in treatment of spasms in gastro-intestinal tract and bile ducts as well as it is used as vasodilator (Mishra *et al.* 2010a). It acts by inhibition of a phosphodiesterase, which is responsible for degradation of cyclic AMP, a common second messenger in the cell (Mishra *et al.* 2010b).

viii) Tropane alkaloids

Tropane alkaloids are alkaloids with nitrogenous bicyclic rings. The tropane class of alkaloids are found mainly in Solanaceae family containing the anticholinergic drugs hyoscyamine and scopolamine. Scopolamine isolated from *Duboisia* and is commonly used as transdermal patch to combat motion sickness (Chan *et al.* 2006; Patel and Ezzo 2009). Cocaine is yet another famous tropane alkaloid. It is isolated from the coca plant and works as a stimulant and appetite suppressant by blocking dopamine reuptake in the brain. Cocaine was actually marketed as a children's toothache medicine in the early 1900s (Dickerson and Janda 2005; Favrod-Coune and Broers 2010).

Atropine, which is isolated from *Atropa belladonna* (deadly nightshade), is another example of tropane alkaloids. Atropine lowers the activity of all the muscles and glands controlled by parasympathetic nervous system, as it is a competitive inhibitor for muscarinic acetylcholine receptors in synapses of parasympathetic nervous system (Takeda *et al.* 2003).

Alkaloids with exocyclic N and aliphatic amines

In their chemical structure, these alkaloids have N outside the cyclic structures (Trease and Evans 1959; Joule and Mills 2000). This category is further subdivided into the following three main categories:

1) Erythrophleum bases

Classification of these alkaloids is based on the appearance of an aminoethanol group. An example of this category of alkaloids is cassaine, which is a cardiotoxic alkaloid that can cause anorexia, defective vision, increased heart sounds, and dyspnea (Phoenix *et al.* 2008).

2) Phenylalkylamines

The main structural component of these alkaloids is an alkylamine attached with two phenyl group (Awasthi and Yadav 2007). This group tends to have hallucinogenic properties and is, therefore, one of the major groups in psychedelic drugs. Most of these psychedelic drugs are used for recreational purposes. For example, mescaline, a phenylalkylamine, is more commonly known as ecstasy. Alkaloids from this category have also been used in order to block voltage-gated Ca channels by binding within the intracellular mouth of ion conducting pore. There are several naturally occurring phenylalkylamines in human body such as dihydroxyphenylalanine (DOPA), dopamine, tyrosine, and adrenaline. Some of these naturally occurring molecules are used as protein building blocks (amino acids) and also for the execution of motive, cognitive, and neuroendocrinic functions. Several of the molecules like dopamine are also involved in pathogenesis of multiple neuropsychiatric diseases such as Parkinson's disease, Alzheimer's disease, and schizophrenia. The Ca channel blocking molecules have antiarrhythmic and antihypertensive properties and can be highly toxic if they bind too strongly to the ion channel preventing the inflow of Ca for an extended period of time (Awasthi and Yadav 2007).

3) Alkaloids having an exocyclic N

Alkaloids like colchicine have an exocyclic N, are normally more biogenetically related to alkaloids with an isoquinoline group. Colchicine itself inhibits the polymerization of

tubulin protein, which is required for mitosis; so it can be used as an antitumor drug, as cancer cells rapidly go through mitosis (Pelletier 1999).

Putrescine, spermidine and spermine alkaloids

Putrescine, spermidine and spermine bases are part of a group of natural products known as biogenic amines; thus, they are not considered as alkaloids (Karovicova and Kohajdova 2005). This class of alkaloids arises because they are derived using these three bases as building blocks. A classic example from the spermidine sub group is Oncinotine, the main alkaloid of *Oncinotis nitida* (Apocynaceae).

Diterpene and steroidal alkaloids

Terpenes are another form of naturally occurring products that are derived from isoprene subunits (Mishra *et al.* 2009). Terpene is a 10-C molecule; thus, diterpenes are two terpenes put together creating twenty C molecules. Although terpenes do not have a basic N base within their isoprene subunits, basic N can be added to the molecule, classifying certain terpenes as alkaloids. The diterpenes can be categorized into the following two major groups:

1) Diterpenes with a 20-C skeleton

These alkaloids belong to several plant families such as Rubunculaceae, Garryaceae, Compositae, and Rosaceae (Mishra *et al.* 2009). The 20-C backbone can be arranged into two major forms:

A) Veatchine skeleton (e.g. veatchine, cuauchichicine, and songorine);

B) Atisine skeleton: This arrangement differs from the veatchine skeleton by having a six-membered ring instead of a five-membered ring, e.g. atisine, atidine, hetisine, ignavine, and kobusine.

2) Diterpenes with a 19-C skeleton

The diterpenes, which have lost a C molecule during their synthesis, belong to this group. They are most commonly found in *Aconitum* and *Delphinium* species (Diaz *et al.* 2005; Forgo *et al.* 2011). Due to the 20-C backbone, diterpene skeletons have two different arrangements:

A) Lycocotonine skeleton, e.g. aconitine, hypaconitine, delphinine, and lycocotonine;

B) Heteratisine skeleton: This arrangement differs from lycocotonine due to addition of an O functional group to produce a lactone.

Peptide alkaloids

These alkaloids are derived from short peptide chains or the chains, which resemble with the peptides. Most of these alkaloids have already been classified under previous categories as many alkaloids are derived from amino acids. The alkaloids considered in this category contain an obvious peptide backbone. For example, celenamide-E is a tripeptide alkaloid in Patagonian sponge, *Cliona chilensis* (Sugumaran and Robinson 2010). This molecule shows antibiotic activity against Gram-negative bacteria.

The chemical classification of alkaloids is universally adapted and depends on the type of heterocyclic ring structure present. It is usual to classify alkaloids according to amino acids (or their derivatives) from which they arise. Thus the most important classes are derived from the amino acids ornithine and lysine; or those derived from aromatic amino acids phenylalanine and tyrosine; or those derived from tryptophan and a moiety of mavelonoid origin. Besides, a number of such compounds are also derived from anthranilic acid or from nicotinic acid.

1) Alkaloids derived from lysine

Such alkaloids, e.g. pelletierine and pseudopelletierine, are found in the bark of pomegranate trees (Khare 2004). Another example is sedamine that is one of the 600 alkaloids isolated from the genus *Sedum*, a common garden plant, while halosaline, a minor constituent, is obtained from *Haloxylon salicornicum* (Cossy *et al.* 2002). Lycopodine is obtained from various species of the moss *Lycopodium*.

The comparison of structures of pelleterine, sedamine, and halosaline suggests that each of these molecules arises from the reaction of a common intermediate with a different Δ -ketoacid. The most likely intermediate is thought to be Δ^1 -piperideine.

2) Alkaloids derived from phenylalanine and tyrosine (benzylisoquinoline)

The simplest members of this group of alkaloids are compounds that contain β -phenylethylamine, $C_6H_5CH_2CH_2NH_2$, skeleton, e.g., morphine, codeine, thebaine, etc. (Ziegler *et al.* 2005).

3) Alkaloids derived from tryptophan

These are quinine type alkaloids derived from tryptophan. Quinine skeleton includes atoms derived from non-amino acid sources, in particular the terpene geraniol. However, feeding experiments, using isotopically labelled geraniol and tryptophan, showed involvement of molecular rearrangements (Dewick 2011).

BIOSYNTHESIS OF ALKALOIDS

Amino acid, the building blocks of alkaloids, are modified by decarboxylation, aldol condensation, reductive amination or methylation. Successive alterations using these relatively few biosynthetic transformations lead to formation of specific alkaloids.

Biosynthesis of terpenoid indole alkaloids

It begins with the conversion of tryptophan to tryptamine and secologenin and ends with the synthesis of terpene indole alkaloids (Facchini 2001).

Biosynthesis of benzylisoquinoline alkaloids (BIA)

The BIA biosynthesis begins with decarboxylations, ortho-hydroxylations, and deaminations of tyrosine that convert it into both dopamine and 4-hydroxy phenylacetaldehyde (Rueffer and Zenk 1987; Facchini 2001).

Biosynthesis of tropane alkaloids

Biosynthesis of tropane alkaloids starts with ornithine and finally ends with the synthesis of tropane alkaloid (Facchini 2001).

Biosynthesis of purine alkaloids

Purine alkaloids, such as caffeine, theobromine, and acrine, are widely distributed in plant kingdom. Recent metabolic studies on tea and coffee have elucidated the biosynthesis of caffeine. The major route begins with xanthosine and proceeds through three N-methylations via 7-methylxanthosine, 7-methylxanthine and theobromine (Suzuki *et al.* 1992; Ashihara *et al.* 1996, 1997) although a number of minor pathways have also been suggested (Kato *et al.* 1996; Schultheiss *et al.* 1996; Facchini 2001).

REGULATORY ROLE OF MINERALS IN ALKALOIDS

An alkaloid is a naturally occurring nitrogenous organic molecule that has pharmacological effect on humans and other animals. Being a nitrogenous compound, availability of N directly influences the level of alkaloids (Demeyer and Dejaegere 1998). Biosynthesis of alkaloids, although controlled genetically, could be affected by different environmental factors, such as light, high temperature, stress and nutrients (Andrijany *et al.* 1998; Chatterjee *et al.* 1988; Lovkov and Buzuk 1998). The mode of action of various groups of alkaloids varies depending on the level of N available. Several decades back, Mothes (1928) conducted a study to look into the effect of different levels of added N on the plant of *Nicotiana* and noticed that initially alkaloid content increased rapidly upon addition of first level of N but additional level of N decreased the total amount of alkaloids. Previously it was noted by geneticist and chemist that type of fertilizer does not affect alkaloids. Later on, Marten *et al.* (1974) studied that ammonium source of N caused greater increase in total alkaloid content than did a nitrate source in Canary grass. However, later on Pavlikova and Tlustos (1994) confirmed the findings of Marten *et al.* (1974). Similar study was also conducted by Yadav *et al.* (1984) on poppy and suggested three important factors, i.e. supply, form and date of application of N in the form of fertilizer, play decisive role in alkaloid production. They suggested that application of N increased the alkaloid content in the strains in which alkaloid content was already high. Similarly in *Catharanthus*, N in NO_3^- form enhanced the alkaloid content up to 50% (Miranda-Ham *et al.* 1996), while application of a mixture of NO_3^- and NH_4^+ decreased the alkaloids by 45%. It was reported in opium poppy that direct dose of N as well as the split doses not only affected the alkaloid concentration but also affected the habits of poppy and seed yield (Kharwara *et al.* 1988; Jain *et al.* 1990; Pavlikova and Tlustos 1994).

Not only N, other macro- and microbiogenous elements also modulate the formation and accumulation of isoquinoline (papaverine) alkaloids in opium poppy (Lovkov *et al.* 2006). In their experiment, they took 12 elements and clustered them into three groups. Group I included molybdenum, cobalt, tungsten, and Cr, group II had Cu and nickel and group III had boron, Fe, vanadium, Mn, zinc and Ca. Group I elements stimulated the formation of isoquinoline alkaloids. The elements of group III inhibited the formation and accumulation of alkaloids at various concentrations. Mo, Cr, and W has probably related to the inhibition of nonspecific phosphatases, which contributed to a more pronounced retention of phosphorylated intermediates during the biosynthesis of amino acids, precursors of alkaloids (tyrosine and phenylalanine) and other intermediate substances.

Co has stimulatory effect on isoquinoline alkaloids by activating the biosynthesis enzymes of the initial stage of aromatic amino acids, including 3-deoxy-D-arabino-heptulosonate-7-phosphate synthase (DAHP synthase, EC 2.5.1.54) and 3-dehydroquinate synthase. Unlike Co-dependent isoform, Mn dependent DAHP synthase is located in the plastids and are characterized by feedback inhibition. The Mn blocks the formation of precursor amino acids by feedback mechanism, which results in the inhibition of isoquinoline accumulation. The difference in the effect of Co and Mn may be related not only to the regulatory activity of DAHP synthase isoforms, but also to the variations in intracellular localization of the isoforms (cytosol and plastids, respectively). Lovkov *et al.* (2006), in an experiment on opium poppy with glycerophosphate, showed the effect of Co. The glycerophosphate specifically inhibits two enzymes in the biosynthesis of aromatic amino acids and Co-dependent DAHP synthase, but does affect the Mn-dependent isoform. Iron (Fe) has inhibitory effect, which provides the evidence for the hypothesis that Co has a role in formation and accumulation of isoquinolines. Fe at various concentrations suppresses different steps of biosynthesis, but does not

have an effect on the growth and formation of seedlings. The antagonistic relationship between Co and Fe suggested that effect of Fe is realized via the displacement and substitution of Co in the enzymes catalyzing the biosynthesis of primary alkaloid precursors (aromatic amino acids).

Another microelement Cu also has stimulatory effect on isoquinoline alkaloids by activating Cu-containing enzyme, polyphenol oxidase located in the plastids and alkaloid accumulating vesicles of latex in the plant cell. This enzyme catalyzes the major early stages of biosynthesis of isoquinoline alkaloids, including tyrosine hydroxylation (with the formation of DOPA), transformation of 4-hydroxyphenyl-acetaldehyde into 3,4-dihydroxyphenyl-acetaldehyde, and conversion of *N*-methylcoclaurine to 3-hydroxy-*N*-methylcoclaurine. Vanadium and Ca is associated with the inhibition of isoquinoline synthesis by inhibiting proton ATPases, which determines the transmembrane electrochemical gradient. Vanadium inhibits the active transport by suppressing the supply of energy required for isoquinoline synthesis. Ca strongly inhibits the biosynthesis of isoquinoline alkaloid of *Papaver somniferum* (Buzuk and Lovkov 1985). Ca modulates the passive transport by stabilizing membranes and decreasing the availability of substrates for the enzymes. Due to high degree of compartmentalization of alkaloid biosynthesis and the decreased active transport, passive transport has a major role in the formation and accumulation of isoquinolines. The element boron stimulates consumption of N, which increases synthesis of various N-containing compounds (e.g. amino acids). The above changes have a positive impact on alkaloid formation in opium poppy. In the poppy capsules, there is positive relationship between morphine and Cd and the negative one between papaverine and zinc and codeine and zinc (Lachman *et al.* 2006). It was also found that atmospheric CO₂ influences the alkaloid content in the case of morphine, codeine nascopine and papaverine. Morphine had maximum response to CO₂ and as a fraction in total alkaloid percentage (Ziska *et al.* 2005, 2008).

In *Catharanthus*, Zn and N play an important role in the synthesis of alkaloids. Zn is indispensable for the synthesis of tryptophan, which is the precursor of indole alkaloids; whereas N is a constituent of the alkaloids. It was reported previously that K had inhibitory role in the synthesis of alkaloids. Depletion of K usually results in an increase in the percentage of alkaloids, while the total percentage of alkaloids is decreased sometimes. In Lupins, K-depleted plants had a higher alkaloid level (Mironenko 1975). Gentry *et al.* (1969) found that P and K together greatly reduced the perloline biosynthesis in tall *Festuca arundinacea* trees, whereas N increased the perloline biosynthesis therein. Other element such as P had significant positive effect on the alkaloid production (Mironenko 1975; Dovrat and Goldschmidt 1978; Chandra 1981), while there was a negative effect in the case of *Trifolium repens* (Jose *et al.* 2000). Ca activates the accumulation of tropane and indole alkaloids in *Hyoscyamus niger* and *Catharanthus roseus*.

In *Datura stramonium*, high amount of Co decreases the yield of all plant parts and alkaloid content of leaves; while in roots, alkaloid content rose with increasing Co rates (Tammaru 1980). However, in *Atropa belladonna*, there was a significant relationship between Co supply and alkaloid content (Petrishek *et al.* 1983). The level of NO₃⁻ decreased the alkaloid content in young plants of *D. stramonium* due to its association with vegetative growth. However, increase in N₂ uptake increases alkaloids in mature plants due to its association with generative parts (Demeyer and Dejaegere 1998). In mature plant, N₂ increased alkaloid content as a result of high metabolic activity and an increase in amino acid. In *Ricinus communis*, the content of high NH₄NO₃ showed an increase in ricine concentration.

In *Colchicum autumnale* there is significantly positive effect of Co and Ca elements on the alkaloid synthesis (Poutaraud and Girardin 2005). A sufficient supply of N, K, Mg, B, Mo, Zn, Fe and Cu increased the galanthamine content in *Leucojum aestivum*.

Tso *et al.* (1973) tested the effect of 54 rare elements on nicotine level of *Nicotiana tabacum* L. Be, Cu, Pd, Pt, and Sm increased the nicotine yield over 25%, whereas Bi, Co, Ho, Pb, Ni, Rb, Ag, Ti, Sn, U V and Zr decreased the nicotine yield. The elements Cs, Er, Li, Rh, Ru, Se, Sr, Ti, and Yb possibly increased the nicotine level less than 25%, whereas As, Ce, Cr, Dy, Gd, I, Mo, Nd, Re, Ta, and Th possibly decreased it. Other elements including Al, Ge, Au, Hf, In, Ir, La, Lu, Hg, Os, Pr, Sc, Te, Tb, Tm, W, and Zn showed no significant effects on nicotine level. This increase in nicotine level in tobacco plants was as a result of the interaction of the elements applied together.

CONCLUSION

Minerals have a diversified role in medicinal plant metabolism. The regulations of metabolic activities remain under strict control of mineral nutrients that are solely provided by soil. Severity or scarcity of minerals causes multifarious effects on plant metabolism. The mineral nutrients are classified as macro and micronutrients depending upon their requirement to the plants. Among different macro elements, N is a major constituent of protoplasm, chlorophyll, amino acids, amides and alkaloids. P is essential to the vital growth processes in the plants, Ca and Mg are the constituents of cell wall, chlorophyll as carrier of P element, K is used in the formation of carbohydrates and proteins and S occur as a constituent of amino acids. Micro elements, required for plant metabolism, include Fe and Mn for chlorophyll formation, boron as a catalyst, and Zn and Fe as a co-factor in some enzymes. Co is required for N fixation in lower plants, while Na as a replacer of K in the regulation of stomatal opening and closing. Though minerals have various regulatory roles in different metabolic processes occurring in plants, the present chapter deals only with their roles in photosynthesis, respiration, protein synthesis, ion uptake and alkaloid biosynthesis, especially in medicinal plants. Both macro and micro elements play major roles in catabolic and anabolic activities in the processes of photosynthesis and respiration, working as a driving force of plant life cycle. Proteins, which are required for structural integrity of plants, are constituted by twenty amino acids; they are also solely dependent on minerals for their structure formation, action and behavior. Different ionic forms of minerals also play regulatory roles in translocation of various solutes and ionic uptake occurring through the biological membranes. Apart from being involved in physiological activities, different minerals also have regulatory roles in alkaloid biosynthesis in different plants. Minerals in the form of fertilizers directly enhance the alkaloid content in most medicinal plants. A number of studies have been carried out regarding various aspects of plant metabolism and interactions of different minerals on various plants, but still a gap in literature exists with respect to specific roles of minerals in medicinal and aromatic plants, which needs further researches to be conducted in depth.

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