

Antioxidant and Polyphenol Oxidase Activity of Some Tunisian Pearl Millet (*Pennisetum glaucum* (L.) R.Br.) Ecotypes

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ABSTRACT

Pearl millet is a rich source of various phytochemicals, including phenolic acids. Natural compounds in food are an important health-protecting factor. In Tunisia, many autochthonous pearl millet ecotypes have generated interest because their nutritional qualities. In order to provide information on the composition of pearl millet grown under local conditions, seven ecotypes were analyzed to evaluate their nutraceutical and antioxidant properties. The nutraceutical properties were determined by evaluating the total phenolic while the antioxidant properties were studied using the DPPH free radical scavenging activity. The results showed that the total phenolic varied widely between ecotypes (from 198 to 323 mg GAE/100 g). The DPPH radical scavenging activity of all the ecotypes was relatively high. The antioxidant activity of pearl millet flour significantly varied among ecotypes and ranged from 62.5 to 86.4%. Fractionation of phenolic extracts by HPLC showed 3 major peaks and several minor peaks: *Trans*-cinnamic (486 to 677 µg/g of extract), protocatechic (127 to 452.4 µg/g of extract) and hydroxybenzoic (253.5 to 437.6 µg/g of extract) were the most important. Identifying ecotypes growing under local agricultural conditions with significant levels of beneficial factors could not only provide health benefit to consumers but also promote the value-added cultivation and stimulate industrial and economic growth.

Keywords: autochthonous, polyphenols, phytochemicals, nutraceutical, Tunisia, value added

INTRODUCTION

Pearl millet (*Pennisetum glaucum* (L.) R. Br) is one of the important crops in semi-arid areas of Africa and India (Andrews and Kumar 1992; O'Kennedy *et al.* 2006). It is a vitally important cereal for the maintenance of food security in Africa and, together with sorghum, represent about half the total cereal production on the continent and as such are a major source of dietary energy and protein for some 1 billion people in the semi-arid tropics (Rooney 2004; Selton and Taylor 2004). Pearl millet proteins are nutritionally important. As they are gluten-free, they are suitable for coeliacs (Kasarda 2001). Interest in bioactive compounds with antioxidant capacity and potential health benefits in Pearl millet is increasing but data on these antioxidants remains scarce in the literature. Pearl millet, like others cereals, in addition to being primary sources of carbohydrates, also provide trace minerals, dietary fibre and bio-active compounds (Madhujith and Shahidi 2006).

Pearl millet is an important source of some minerals particularly iron and zinc (Serna-Saldivar and Rooney 1995; Adeola and Orban 1995). It has high levels of lipids, high quality and well-balanced proteins (Elyas and El Tinay Yousif 2002). Pearl millet has health-promoting properties, in particular its antioxidant activity (Chung and Pomeranz 1985; Madhujith and Shahidi 2007). It is used as a nutraceutical and as functional food (Dykes and Rooney 2006). Pearl millet has anti-carcinogenic properties (Van Rensburg 1981; Chen *et al.* 1993). It is also a potentially important source of polyphenol and cholesterol-lowering waxes (Hwang *et al.* 2002). It could serve as an important source of phytochemicals (Awika *et al.* 2003).

Polyphenol compounds can affect a wide range of cell biological functions by virtue of their radical scavenging

properties (Aruoma 1993; Rakic *et al.* 2006). Many studies have suggested the role of phenolic compounds as the major source of natural antioxidants in foods of plant origin (Hagerman *et al.* 1998; Balasundram *et al.* 2006). Several workers have highlighted the relationships between food phenol composition and antioxidant activity. While some authors found a correlation between polyphenol content and antioxidant activity, others found no relationship (Moure *et al.* 2001; Heim *et al.* 2002).

Despite the high levels and diversity of phytochemicals in pearl millet, research on this crop as a source of valuable health promoting compounds lags behind other commodities (e.g., fruits and vegetables) and still under researched compared to other cereals. As a result, utilization of pearl millet fractions in foods to improve nutrition is very limited.

Several studies demonstrated that fortification of diet with food components rich in phenolic acids has been shown to impart antimutagenic, antiglycemic and antioxidative properties, and this has been exploited for the development of healthy food formulations (Friedman 1997).

Pearl millet has a big potential, given its agronomic properties, as well as the emerging evidence on the biological effects of the phytochemicals present in the grain.

In Tunisia, pearl millet isn't the staple food of rural populations as in the other countries of Africa. Nevertheless, it occupies a very important part of surfaces every year in the centre and in the South of the country (FAO 2003). All pearl millet production is used for a variety of food products. Many autochthonous pearl millet ecotypes have generated interest in Tunisia because their nutritional qualities. However, little information is currently available.

Identifying ecotypes growing under local agricultural conditions with significant levels of beneficial factors could not only provide health benefit to consumers but also pro-

Table 1 Pearl millet ecotypes surveyed, and their ecological parameters.

Ecotype	Origin	Bio-climatical strata	Annual rainfall (mm)	Annual average temperature (°C)	January average temperature (°C)	July average temperature (°C)
SM	Tataouine	Saharan (Sup)	25-100	22.3	10.5	29.3
KS	Kairouan Kairouan	Arid (Med)	200-400	19.9	11.5	28.7
D - MD- BG	Médnine	Arid (Inf)	100-200	21.3	12.4	26.9
A	Ariana	Semi-arid	400-600	18.3	12.1	26.0
HZ	Mehdia	Arid (Sup)	200-400	19.5	11.9	26.3

Data according to the National Institute of Meteorology (INM), 2009, Tunisia.

mote the value-added cultivation and stimulate industrial and economic growth. Biochemical properties could offer reliable tools to distinguish cultivars or ecotypes and may be considered as criteria of selection for specific food applications. The use of pearl millets not only provides farmers with a market for their products but also saves foreign exchange, which would otherwise be required to import cereals. In fact, food manufacturers as consumers have begun to move towards functional foods with special health effects (Javanmardi *et al.* 2003). Actual studies have focused on genetic improvement of sorghum for biofuel production (Liang 2012) and on genetic variability (iron and zinc content) in diverse sorghum germplasm (Ashok Kumar *et al.* 2012). However, studies regarding antioxidant and nutritional composition are still scarce (Awika *et al.* 2003; Viswanath *et al.* 2009).

In order to provide information on the composition of pearl millet grown under local conditions, seven ecotypes were analyzed to evaluate their phenolic compounds and antioxidant properties.

MATERIALS AND METHODS

Plant material

Seven autochthonous pearl millet ecotypes (*Pennisetum glaucum* (L.) R. Br) from different ecological regions from Tunisia were collected (Table 1). Each ecotype is affected of an abbreviation symbolizing the zone of origin.

Plant growth and treatments

Collected grains from each ecotype were sowed at the farm of the Tunisian Agricultural Research Institute in Tunis during the 2007 cropping season. The site is located at 36° 51' latitude and 10° 11' longitude. The soil of the experimental site was clay loam. They were sown on 22 May 2007 into randomized block design with four replications. After fructification of the cultivated plants, the grains were homogeneously collected for a biochemical investigation. In order to keep relatively uniform sample sizes, we limited our random sampling to 20 plants per ecotype. Equal amounts of grains for each ecotype were pooled and ground to a fine powder to obtain the flour.

Chemicals

Gallic, ferulic, ellagic, vanillic and syringic acids were obtained from Sigma-Aldrich, Chemical Co., Milan. Methanol was of analytical grade and purchased from Carlo Erba (Milan, Italy).

Preparation of polyphenol extracts

About 1 g of flour fractions were refluxed with 1% HCl in CH₃OH (100 ml) for 30 min at 90°C. The extraction was repeated with the residue and the extracts were pooled, neutralised with NaOH, centrifuged (10 min, 3000 rpm), concentrated in a rotary flash evaporator, freeze-dried and stored at 20°C till use (Viswanath *et al.* 2009).

Determination of total phenolic contents (TPC)

Total phenolic compounds from the concentrated extract were determined by the Folin-Ciocalteu micro method (Linkard and

Singleton 1977). About 20 µl of extracts solution were mixed with 1.16 ml distilled water and 100 µl of Folin-Ciocalteu reagent, followed after 1 min and before 8 min by 200 µl of Na₂CO₃ solution (20%). Subsequently, the mixture was incubated in a shaking incubator at 40°C for 30 min and its absorbance was measured at 760 nm. Gallic acid (0.1 mg/ml) was used as the standard and the phenolic content in the extract/fraction was expressed as milligram equivalent of gallic acid (GAE) per 100 g flour.

Radical scavenging activity

The antioxidant activity of native and processed little millet extracts were also measured by the DPPH radical scavenging method (De Ancos *et al.* 2002). An aliquot (10 µl) of acidified methanolic extract was mixed with distilled water (90 µl) and 3.9 ml of methanolic 0.1 mM DPPH solution. The mixture was thoroughly vortex-mixed and kept in dark for 30 min. The absorbance was measured at 515 nm. The result was expressed as percentage of inhibition of the DPPH radical. The percentage of inhibition of the DPPH radical was calculated according to the following equation:

$$\% \text{ inhibition of DPPH} = [\text{Abs control} - \text{Abs sample} / \text{Abs control}] \times 100$$

where Abs control is the absorbance of the DPPH solution without the extracts.

Separation of phenolic compounds

Separation and identification of the polyphenolic components in all extracts was carried out by HPLC (Shimadzu, LC10 A), on a reversed phase chromatography on silica based C18-bonded phase columns (25 cm - 4.6 mm), using a diode array detector operating at different wavelengths (290, 295 and 300 nm). A solvent system comprising of water/methanol (70:30) was used as mobile phase at a flow rate of 1 ml/min. About 100 mg each of freeze dried extracts were dissolved in 1 ml of 50% methanol, 20 µl of each sample was injected into the column. Standards such as gallic, ferulic, ellagic, vanillic and syringic acids were used for identification and quantification of phenolic acids in the samples. Quantification of phenolic acids was carried out by measuring the area under respective peaks and plotting against a standard graph prepared for each individual phenolic acid used (Onyeneho and Hettiarachchy 1992; Rodríguez-Delgado *et al.* 2002).

Statistical analysis

Quantitative presented data are mean ± standard deviation of triplicate tests. Each experiment was performed in triplicate, and the results were expressed as mean values ± standard deviation. Statistical significance was determined by one-way analysis of variance (ANOVA) followed by a multiple comparison test (Tukey's test) at 5% level of significance. Values with $P < 0.05$ were considered statistically significant.

RESULTS AND DISCUSSION

The antioxidative phytochemicals in grains, vegetables and fruits have received increased attention recently for their potential role in prevention of human diseases as well as in food quality improvement (Chu *et al.* 2002; Nandita 2002; Nandita and Rajini 2004). Phenolics are considered as a major group of compounds that contribute to the antioxi-

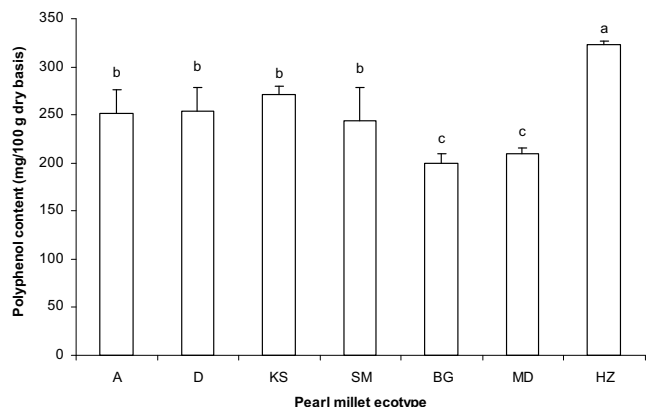


Fig. 1 Total polyphenol content of Tunisian pearl millet ecotypes (A: Ariana, D: Médnine, KS: Kairouan, SM: Tataouine, BG: Médnine, MD: Médnine, HZ: Mehdiá). Data are means of three replicates \pm standard error (Tukey's test, $P < 0.05$).

dant activities of grains. Pearl millet is an important food grains reported to contain significant quantities of phenolic compounds.

Total phenolic content

Total phenolic content (TPC) in whole pearl millet ecotypes varied significantly among ecotypes (Fig. 1). The highest TPC was observed in HZ ecotype with 323 mg/100 g (mg equivalent of gallic acid (GAE)/100 g) while BG showed the lowest TPC (198 mg GAE/100 g).

Phenolic acids are presents in cereal grains (Andlauer and Furst 1998; Towwo *et al.* 2003; Viswanath *et al.* 2009). Their content in cereals is usually less than 1% of dry matter, except for some sorghum cultivars (Subba Rao and Muralikrishna 2002) and finger millet varieties (Chethan and Malleshi 2007a).

The TPC of whole flour from autochthonous pearl millet varied significantly among the ecotypes but it is fairly low compared to many millet varieties cultivated which have polyphenol content of approximately 2.6% (Viswanath *et al.* 2009). Some authors (Sehgal and Kawatra 1998) reported TPC of Bajra ecotype (HHB-50) to be 764 mg/100 g.

Total polyphenols content were 319 and 294 mg/100 g for Composite Pop III and Baladi 1997, respectively (Elyas and El Tinay Youssif 2002). They demonstrated that this in agreement with the fact that polyphenols content of pearl millet is fairly high, as reported by (Alka-Sharma and Kapoor 1996).

Polyphenol quantity and quality in plant foods can vary significantly according to different intrinsic and extrinsic factors such as plant genetics and cultivar, soil composition and growing conditions, maturity state and post harvest conditions, among others (Jaffery *et al.* 2003).

Table 2 Antioxidant properties of pearl millet ecotypes.

Ecotype	% DPPH inhibition
A	70.0 b
D	71.7 b
KS	72.0 b
SM	74.2 b
BG	62.5 c
MD	97.2 c
HZ	96.4 a

Values with same letters (a,b,c,d,e within columns) are not significantly different (Tukey's test, $P < 0.05$).

Antioxidant activity

The DPPH is a stable free radical widely used to determine the antioxidant properties or radical scavenging activity. The radical scavenging capacity against DPPH of all the ecotypes was found relatively high. The antioxidant activity of pearl millet flour significantly varied among ecotypes and ranged from 62.5 to 86.4%. Pearl millet ecotype (HZ) showed the highest antioxidant activity while (BG) ecotype exhibited the lowest (Table 2). These results may be due to the presence of highest TPC in the HZ ecotype since the %DPPH inhibition is directly correlated with TPC (Alothman *et al.* 2009). Significant positive correlations between these two parameters were also reported in numerous analytical studies (Dykes *et al.* 2005; Lu *et al.* 2007).

Fractionation of phenolics

The HPLC profile revealed the complex nature of millet polyphenols, and the phenolic constituents were identified using known standards. The phenolic acids reported in pearl millet ecotypes are listed in Table 3.

Pearl millet ecotypes vary widely in their phenolic composition and content, with both genetics and environment affecting the kind and level of phenolic compounds.

The HPLC profiles indicated 3 major peaks and several minor peaks revealing the complex nature of pearl millet ecotypes. *Trans*-cinnamic, protocatechic and hydroxybenzoic are the most abundant. Gallic, ferulic, coumaric, vanillic and others acids were identified and the other components remain to be identified. Further studies are required to characterise these phenolics.

Cereals contain a wide range of phenolic compounds including benzoic and cinammic acids, anthocyanidins, quinones, flavonols, chalcones, flavanones and amino phenolics (Andreasen *et al.* 2000; Adom and Liuy 2002).

The most important phenolic acids in autochthonous pearl millet ecotypes are respectively: *trans*-cinnamic, protocatechic and hydroxybenzoic for all ecotypes. This result is corroborated by (Hahn *et al.* 1983; Waniska and Bandyopadhyay 1989). They found that the phenolic acids in millets are benzoic or cinnamic acid derivatives.

All millets contain phenolic acids, which are located in the pericarp, testa, aleurone layer, and endosperm (Dykes and Rooney 2006). McDonough and Rooney (2000) repor-

Table 3 Phenolic acid composition of different pearl millet ecotype ($\mu\text{g/g}$ of extract).

Phenolic acid	KS	A	BG	HZ	SM	D	MD
Gallic acid	9.8 a	10.9 a	13.6 a	10.1a	11.2 a	7.3 b	9.4 a
Catechin	3.9 b	4.3 b	23.2 a	3.6 b	18.9 a	0.3 d	1.2 c
Ferulic acid	ND	23.6 b	30.2 a	ND	20.3 b	ND	ND
p-Coumaric acid	0.9 a	ND	0.6 a	1.3 a	1.1 a	0.7 a	0.9 a
Gentisic acid	35.4 a	ND	ND	23.3 b	ND	17.3 b	19.7 b
<i>Trans</i> -cinnamic acid	731.2 a	575.8 d	486.3	677.1 b	513.0 e	643.0 c	633.0 c
Protocatechic acid	452.4 a	365.8 c	127.6 d	414.0 b	182.0 d	433.0 b	387.1 c
Hydroxybenzoic acid	437.6 a	253.5 d	ND	391.0 b	ND	382.0 b	352.0 c
Quercitrin	0.7 bc	2.1 a	0.9 b	1.2 b	1.4 b	0.7 c	0.3 d
Quercitin	1.5 b	2.4 a	1.2 b	0.9 c	2.1 a	1.1 b	1.7 b
Vanillic acid	ND	48.3	ND	ND	ND	ND	ND
Caffeic acid	1.3 a	ND	ND	1.7 a	ND	0.9 b	1.3 a

ND = not detected ; Tukey's test, $P < 0.05$

ted the phenolic acid composition of some millet. In 'Finger' and 'Pearl' varieties, ferulic acid was the highest among all the phenolic acids with 387 and 679.7 mg/100 g dry weight basis. However, in 'foxatil' coumaric acid was the highest with 2133.7 mg/100 g dry weight basis. Phenolic acids consist of two classes: hydroxybenzoic and hydroxycinnamic acids. Hydroxybenzoic acids are directly derived from benzoic acid and include gallic, hydroxybenzoic, vanillic, syringic, and protocatechuic acids, among others. The hydroxycinnamic acids include coumaric, caffeic, ferulic, and sinapic acids (Hahn *et al.* 1984; McDonough *et al.* 1986). Phenolic acids such as caffeic, coumaric, ferulic and proto-catechuic acids are also shown to exert an antifungal effect (Aranowski *et al.* 1980).

CONCLUSIONS

Pearl millet is a tropical cereal that has great diversity in its content of phenolic compounds as well as antioxidant activity. These substances are effective biochemical determinants for pearl millet use as food or as source of bioactive components. Research on bioactive components such as phenolic compounds will contribute to unleash the capacity of pearl millet to be a source of nutraceuticals in food formulations and to help to food security in Africa as well as in many developing countries.

REFERENCES

- Adeola O, Orban JI (1995) Chemical composition and nutrient digestibility of pearl millet (*Pennisetum glaucum*) fed to growing pigs. *Journal of Science* **22**, 177-184
- Adom KK, Liu RH (2002) Antioxidant activity of grains. *Journal of Agricultural and Food Chemistry* **50**, 6182-6187
- Alka-Sharma X Kapoor AC (1996) Levels of antinutritional factors in pearl millet as affected by processing treatment and various types of fermentation. *Plant Foods for Human Nutrition* **49**, 241-252
- Allothman M, Bhat R, Karim AA (2009) Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. *Food Chemistry* **115**, 785-788
- Andlauer W, Furst P (1998) Antioxidative power of phytochemicals with special reference to cereals. *Cereals Foods World* **43**, 356-360
- Andreasen MF, Christensen LP, Meyer AS, Hansen AS (2000) Contents of phenolic acids and ferulic acid dehydromers in 17 rye (*Secale cereale* L.) varieties. *Journal of Agricultural and Food Chemistry* **48**, 2837-2842
- Andrews DJ, Kumar KA (1992) Pearl millet for food, feed and forage. *Advances in Agronomy* **48**, 89-139
- Aruoma OI (1998) Free radicals, oxidative stress and antioxidants in human health and disease. *Journal of American Oil Chemists' Society* **75**, 199-212
- Ashok Kumar A, Belum VS, Reddy B, Ramaiah HK, Sahrawat L, Wolfgang HP (2012) Genetic variability and character association for grain iron and zinc contents in sorghum germplasm accession and commercial cultivars. In: Dobrąnski J (Ed) Sorghum. *The European Journal of Plant Science and Biotechnology* **6** (Special Issue 1), 66-70
- Awika JM, Rooney LW, Wu X, Prior RL, Cisneros-Zevallos L (2003) Screening methods to measure antioxidant activity of sorghum (*Sorghum bicolor*) and sorghum products. *Journal of Agricultural and Food Chemistry* **51**, 6657-6662
- Balasundram N, Sundram K, Sammar S (2006) Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry* **68**, 191-203
- Baranowski JD, Davidson PM, Nagel CW, Brannen RL (1980) Inhibition of *Saccharomyces cerevisiae* by naturally occurring hydroxy cinnamates. *Journal of Food Science* **45**, 592-594
- Belton PS, Taylor JRN (2004) Sorghum and millets: Protein sources for Africa. *Trends in Food Science and Technology* **15**, 94-98
- Chen F, Cole P, Mi ZB, Xing LY (1993) Corn and wheat-flour consumption and mortality from esophageal cancer in Shanxi, China. *International Journal of Cancer* **53**, 902-906
- Chethan S, Malleshi NG (2007) Finger millet polyphenols: Characterization and their nutraceutical potential. *American Journal of Food Technology* **2**, 582-592
- Chethan S, Malleshi NG (2007) Finger millet polyphenols: Optimization of extraction and the effect of pH on their stability. *Food Chemistry* **105**, 862-870
- Chowdari KV, Davierwala AP, Gupta VS, Ranjekar PK, Govila OP (1998) Genotype identification and assessment of genetic relationships in pearl millet [*Pennisetum glaucum* (L.) R. Br.] using microsatellites and RAPDs. *Theoretical and Applied Genetics* **97**, 154-162
- Chu YF, Sun J, Wu X, Liu H (2002) Antioxidant and antiproliferative activities of common vegetables. *Journal of Agricultural and Food Chemistry* **50**, 6910-6916
- Chung OK, Pomeranz V (1985) Amino acids in cereal proteins fractions. In: Finely JW, Hopkins DT (Eds) *Digestibility and Amino Acid Availability in Cereals and Oil Seeds*, American Association of Cereal Chemists, St. Paul, MN, pp 65-107
- De Ancos B, Sgroppo S, Plaza L, Cano MP (2002) Possible nutritional and health-related value promotion in orange juice preserved by high-pressure treatment. *Journal of the Science of Food and Agriculture* **82**, 790-796
- Dykes L, Rooney LW (2006) Sorghum and millet phenols and antioxidants. *Journal of Cereal Science* **44**, 236-251
- Elyas SHA, El Tinay AH, Yousif NE, Elsheikh EAE (2002) Effect of natural fermentation on nutritive value and *in vitro* protein digestibility of pearl millet. *Food Chemistry* **78**, 75-79
- FAO (2003) Statistics series: La production et la surface du mil dans certains pays et dans le monde. *Rome* **57** (177), 1-10
- Friedman M (1997) Chemistry, biochemistry, and dietary role of potato polyphenols - A review. *Journal of Agricultural and Food Chemistry* **45**, 1523-1540
- Hagerman AE, Riedl KM, Jones GA, Sovik KN, Ritchard NT, Hartzfeld PW, Riechel TL (1998) High molecular weight plant phenolics (tannins) as biological antioxidants. *Journal of Agricultural and Food Chemistry* **46**, 1887-1892
- Hahn DH, Faubion JM, Rooney LW (1983) Sorghum phenolic acids, their performance liquid chromatography separation and their relation to fungal resistance. *Cereal Chemistry* **60**, 255-259
- Hahn DH, Rooney LW, Earp CF (1984) Tannins and phenols of sorghum. *Cereal Foods World* **29**, 776-779
- Heim KE, Tagliaferro AR, Bobilya DJ (2002) Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships. *Journal of Nutritional Biochemistry* **13**, 572-584
- Hwang KT, Cuppett SL, Weller CL, Hanna MA (2002) HPLC of grain sorghum wax classes highlighting separation of aldehydes from wax esters and steryl esters. *Journal of Separation Science* **25**, 619-623
- Jaffery EH, Brown AF, Kurilich AC, Keek AS, Matusheski N, Klein BP (2003) Variation in content of bioactive components in broccoli. *Journal of Food Composition and Analysis* **16**, 323-330
- Javanmardi J, Stushnoff L, Locke E, Vivanco JM (2003) Antioxidant activity and total phenolic content of Iranian *Ocimum* accessions. *Food Chemistry* **83**, 547-550
- Kasarda DD (2001) Grains in relation to celiac disease. *Cereal Foods World* **46**, 209-210
- Liang Y (2012) Sorghum: genetic improvement for biofuel. In: Dobrąnski J (Ed) Sorghum. *The European Journal of Plant Science and Biotechnology* **6** (Special Issue 1), 1-9
- Madhujith T, Shahidi F (2006) Antioxidant properties of barley pearled fractions. *Journal of Agriculture and Food Chemistry* **54**, 3283-3289
- Madhujith T, Shahidi F (2007) Antioxidative and antiproliferative properties of selected barley (*Hordeum vulgare* L.) cultivars and their potential for inhibition of low density lipoprotein (LDL) cholesterol oxidation. *Journal of Agriculture and Food Chemistry* **55**, 5018-5024
- Maqbool SB, Devi P, Sticklen M (2001) Biotechnology: genetic improvement of sorghum (*Sorghum bicolor* (L.) Moench). *In Vitro Cell Developmental Biology - Plant* **37**, 504-515
- McDonough CM, Rooney LW, Earp CF (1986) Structural characteristics of *Eleusine coracana* (finger millet) using scanning electron and fluorescence microscopy. *Food Microstructure* **5**, 247-256
- McDonough CM, Rooney LW (2000) The millets. In: Kulp K, Ponte Jr. JG (Eds) *Handbook of Cereal Science and Technology*, Marcel Dekker, Inc., New York, pp 177-201
- Moure A, Cruz JM, Franco D, Domínguez M, Sineiro J, Domínguez H, Nunez MJ, Parajó JC (2001) Natural antioxidants from residual sources. *Food Chemistry* **72**, 145-171
- Nandita S (2002) Studies on the biological activity of potato waste (peel) components for their possible applications. PhD thesis, University of Mysore, Mysore, India, 258 pp
- Nandita S, Rajini PS (2004) Free radical scavenging activity of an aqueous extract of potato peel. *Food Chemistry* **85**, 611-616
- Nithya KS, Ramachandramurthy B, Krishnamoorthy VV (2007) Effect of processing methods on nutritional and anti-nutritional qualities of hybrid (COHCU-8) and traditional (CO7) pearl millet varieties of India. *Journal of Biological Sciences* **7**, 643-647
- O'Kennedy MM, Grootboom A, Shewry PR (2006) Harnessing sorghum and millet biotechnology for food and health. *Journal of Cereal Science* **44**, 224-235
- Onyeneho SN, Hettiarachchy NS (1992) Antioxidant activity of durum wheat bran. *Journal of Agricultural Food Chemistry* **40**, 1496-1500
- Radhouane L (2008) Autochthonous pearl millet ecotype (*Pennisetum glaucum* (L.) R.Br.) response to different sowing dates in Tunisia. *Sjemenarstvo (Seed Science Technology)* **25**, 123-138
- Rakic S, Povrenovic D, Tešević V, Simic M, Maletic R (2006) Oak acorn, polyphenols and antioxidant activity in functional food. *Journal of Food Engineering* **74**, 416-423

- Rodríguez-Delgado MA, González-Hernández G, Conde-González JE, Pérez-Trujillo JP** (2002) Principal component analysis of the polyphenol content in young red wines. *Food Chemistry* **78**, 523-532
- Rooney LW** (2004) Sorghum improvement - Integrating traditional and new technology to produce improved genotypes. *Advances in Agronomy* **83**, 37-109
- Sehgal AS, Kawatra A** (1998) Reduction of polyphenol and phytic acid content of pearl millet grains by malting and blanching. *Plant Foods for Human Nutrition* **53**, 93-98
- Serna-Saldivar S, Rooney LW** (1995) Structure and chemistry of sorghum and millets. In: Dedy DAV (Ed) *Sorghum and Millets: Chemistry and Technology*, American Association of Cereal Chemists, St. Paul, MN, USA, pp 69-124
- Sharma HC, Sharma KK, Seetharama N, Ortiz R** (2000) Prospects for using transgenic resistance to insects in crop improvement. *Electronic Journal of Biotechnology* **3**, 76-95
- Slinkard K, Singleton VL** (1977) Total phenol analysis: Automation and comparison with manual methods. *American Journal of Oenology and Viticulture* **28**, 49-55
- Subba Rao MVSST, Muralikrishna G** (2002) Evaluation of the antioxidant properties of free and bound phenolic acids from native and malted finger millet (ragi, *E. coracana* Indaf-15). *Journal of Agricultural and Food Chemistry* **50**, 889-892
- Towo EE, Svanberg U, Ndossi GD** (2003) Effect of grain pre-treatment on different extractable phenolic groups in cereals and legumes commonly consumed in Tanzania. *Journal of the Science of Food and Agriculture* **83**, 980-986
- Van Rensburg SJ** (1981) Epidemiological and dietary evidence for a specific nutritional predisposition to esophageal cancer. *Journal of the National Cancer Institute* **67**, 243-251
- Viswanath V, Urooj A, Malleshi NG** (2009) Evaluation of antioxidant and antimicrobial properties of finger millet polyphenols (*Eleusine coracana*). *Food Chemistry* **114**, 340-346
- Waniska RD, Poe JH, Bandyopadhyay R** (1989) Effects of growth conditions on grain molding and phenols in sorghum caryopsis. *Journal of Cereal Science* **10**, 217-225